

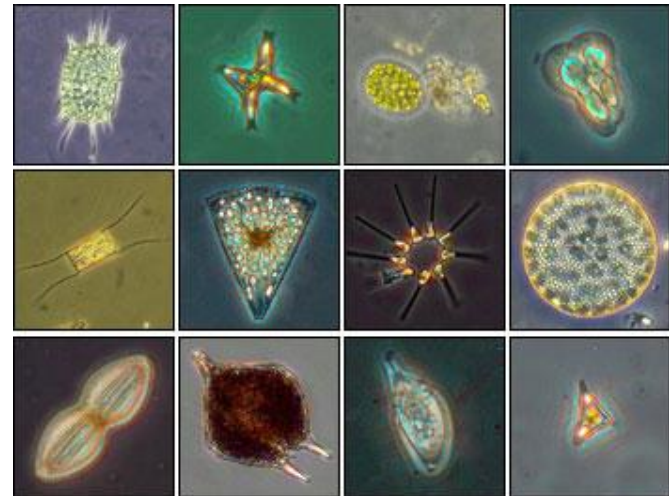
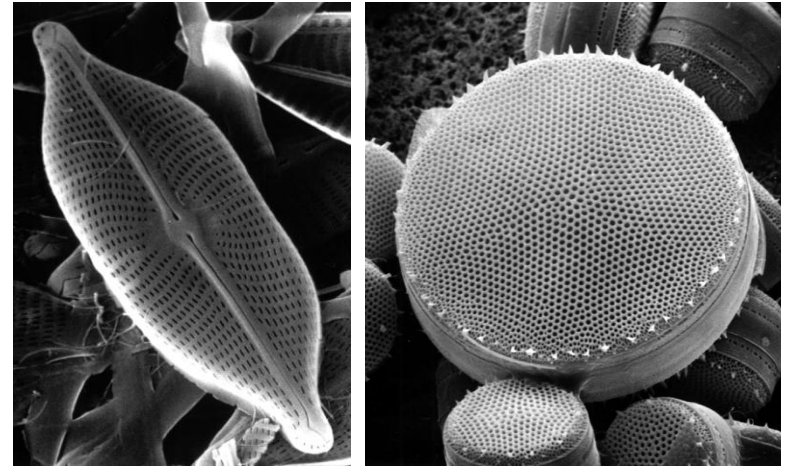
**EBS 566/555 3/3/2010**

Lecture 15

Blooms, primary production  
and types of phytoplankton  
(Ch 2, 3)

## Topics

- Finish blooms
- Major taxa of phytoplankton
- How to measure phytoplankton
- Primary production

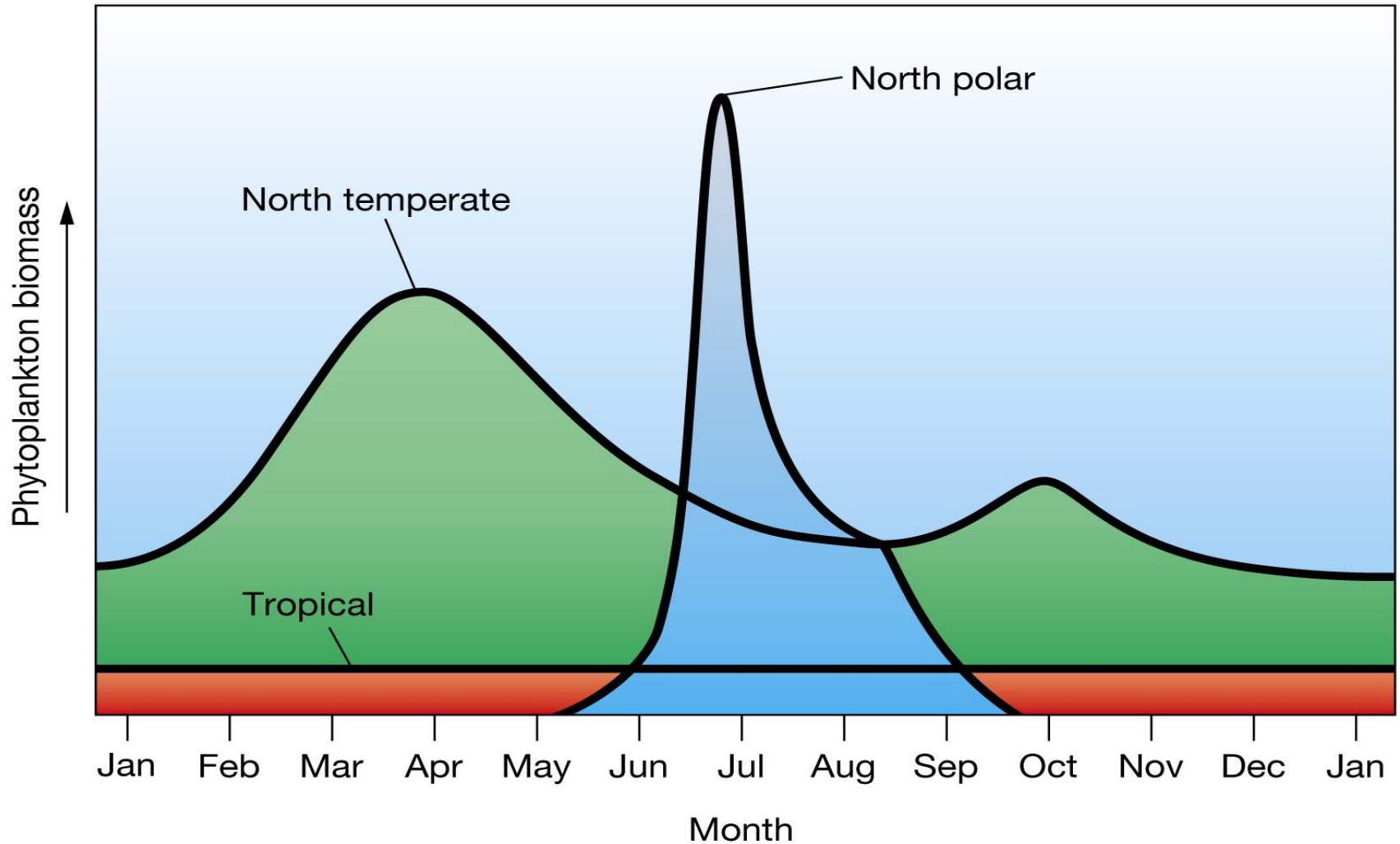


<http://cmore.soest.hawaii.edu>

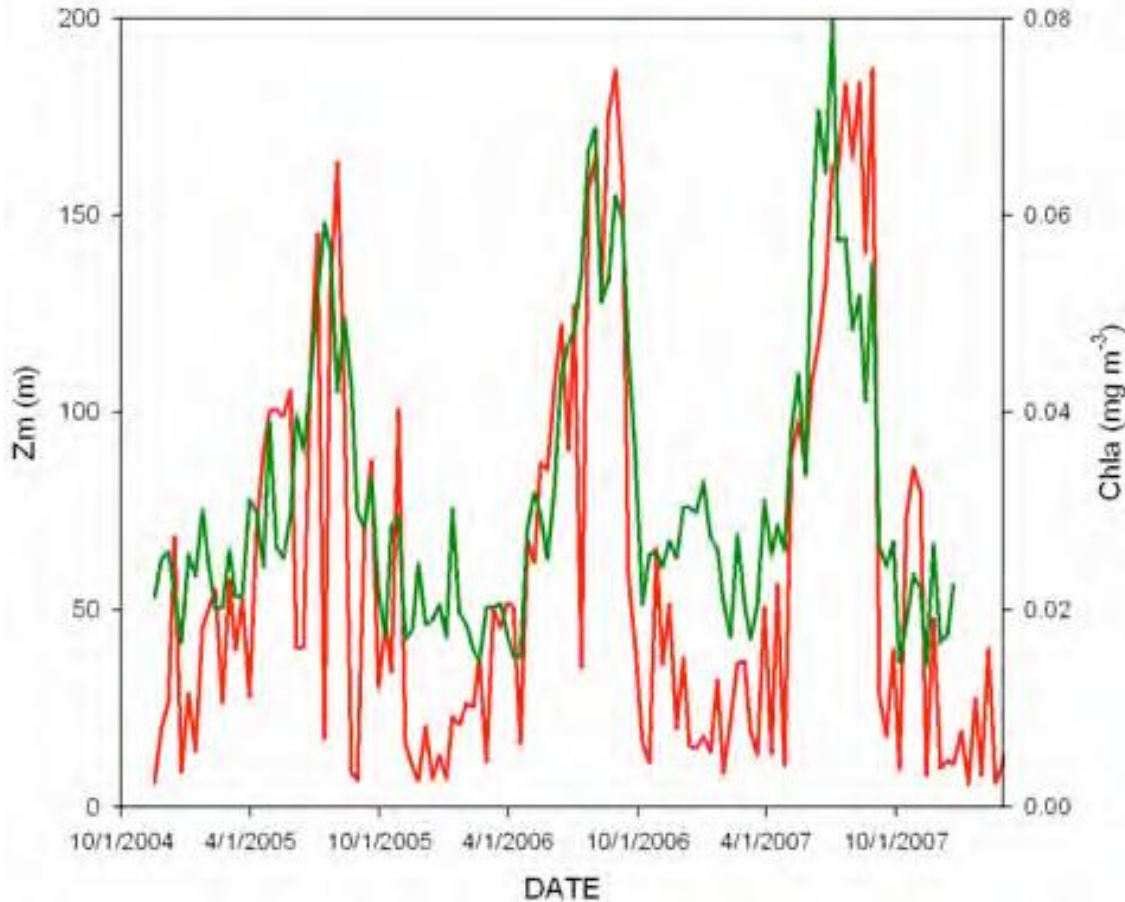
# Alternatives to the Spring Bloom

- Polar and tropical seas
- Shallow water case: stratification not important for bloom initiation, only light is important
- Estuaries (Cloern and Jassby 2008, 2009)
- Extremely oligotrophic regions
- Iron limitation (e.g. subarctic North Pacific)
  - But also applies to spring bloom!
- Grazing regime
- Upwelling systems (coastal, equatorial)

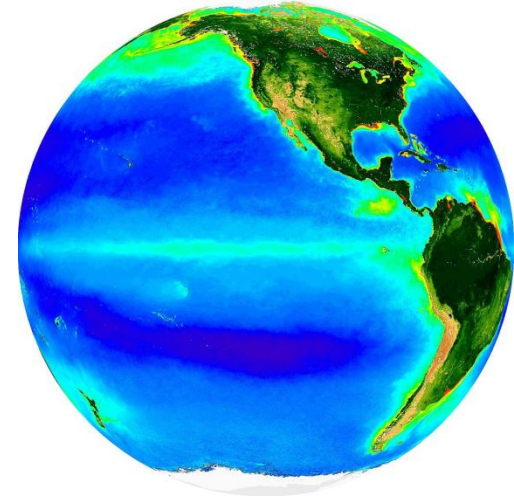
# Global comparison of phytoplankton biomass



# In South Pacific Gyre, chl *a* is highest in winter compared to summer

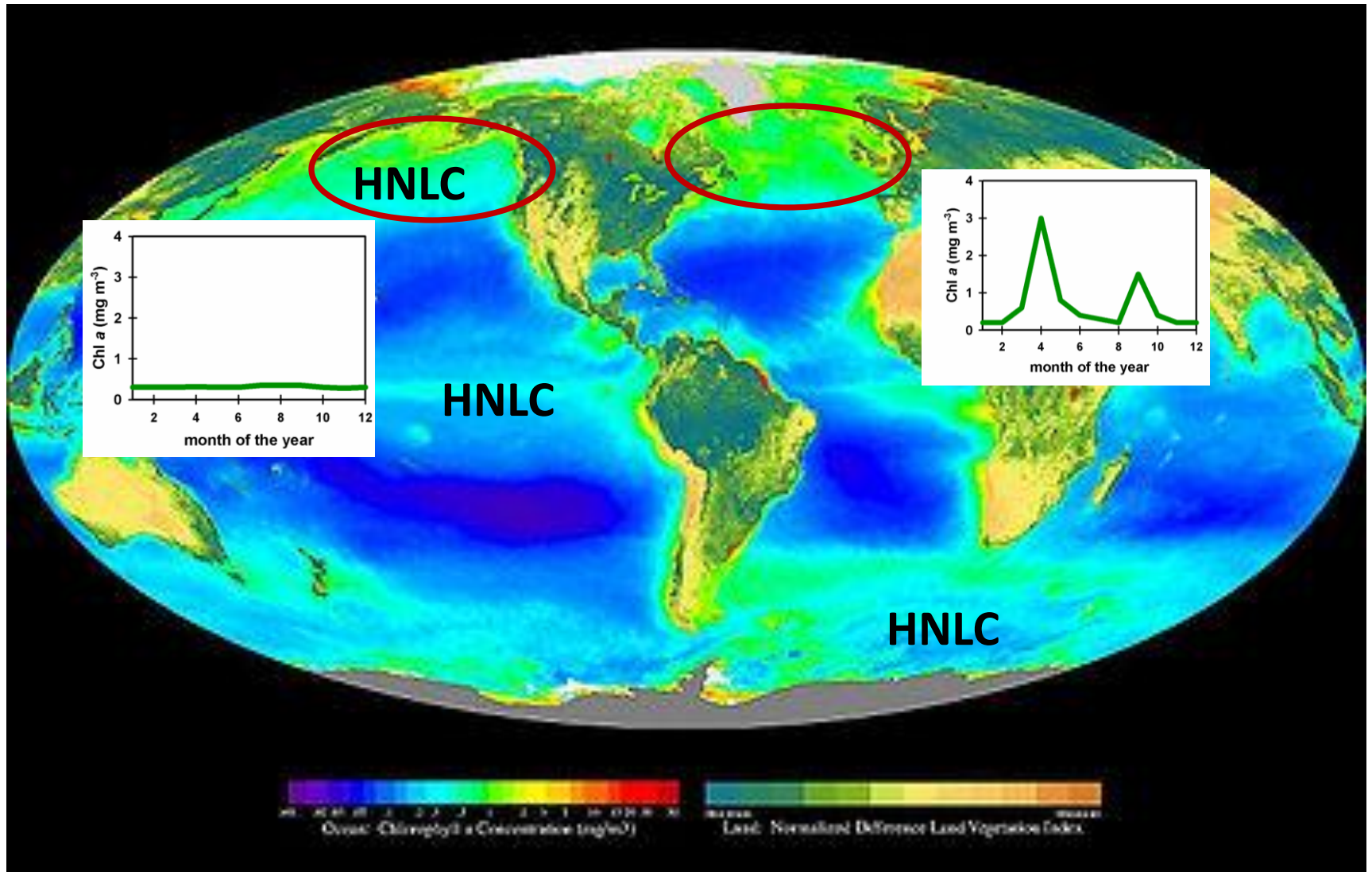


Green = chl *a*  
Red = mixed layer  
depth (determined  
from Argo float)



Claustre, unpubl.

# Annually averaged chl *a*

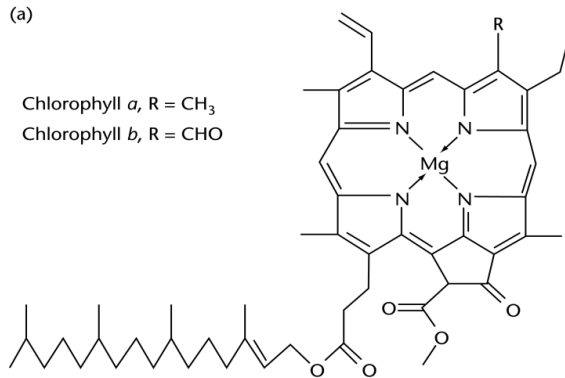


# Fate of bloom phytoplankton

- Shallow decomposition (→ microbial loop, regeneration)
- Export
  - To deep ocean
  - Burial in sediments (→ sedimentary record)
- Grazing
  - Mesozooplankton
  - Microzooplankton
  - Excretion → regenerated nutrients



# We can estimate algal biomass using the photosynthetic pigment, chlorophyll *a*



Plant Cell Chloroplast Structure

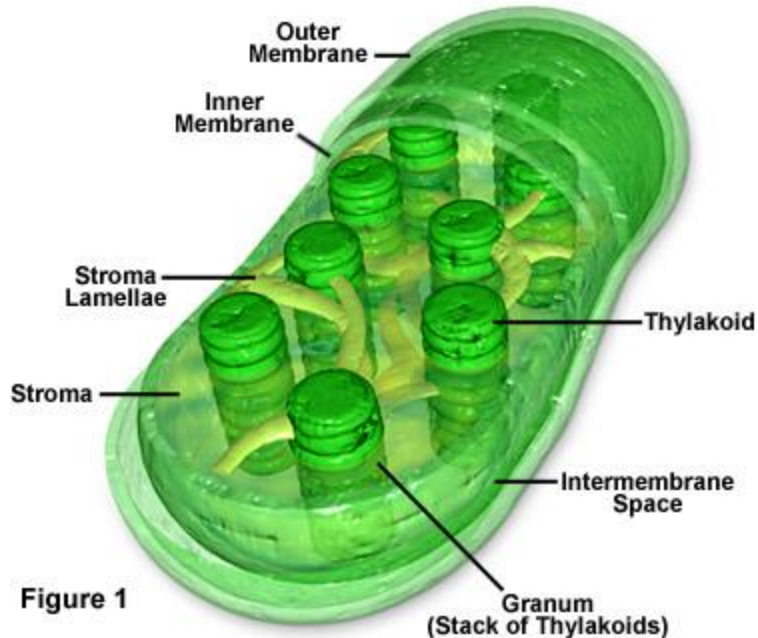
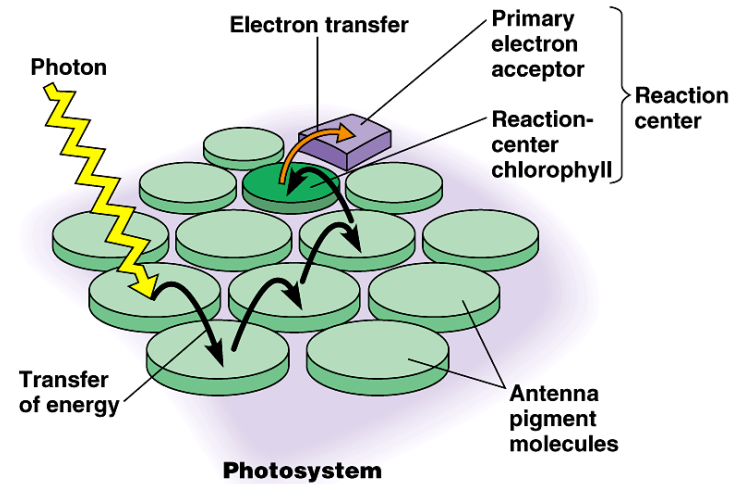
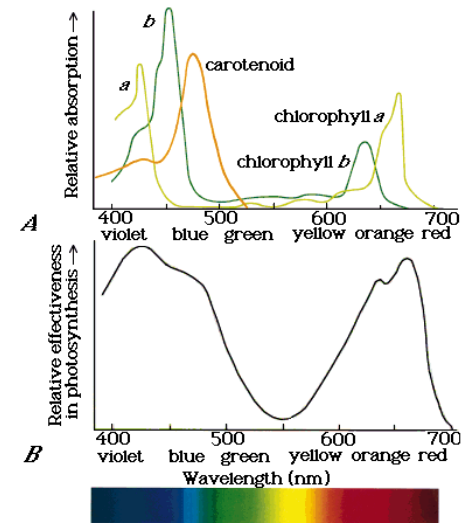


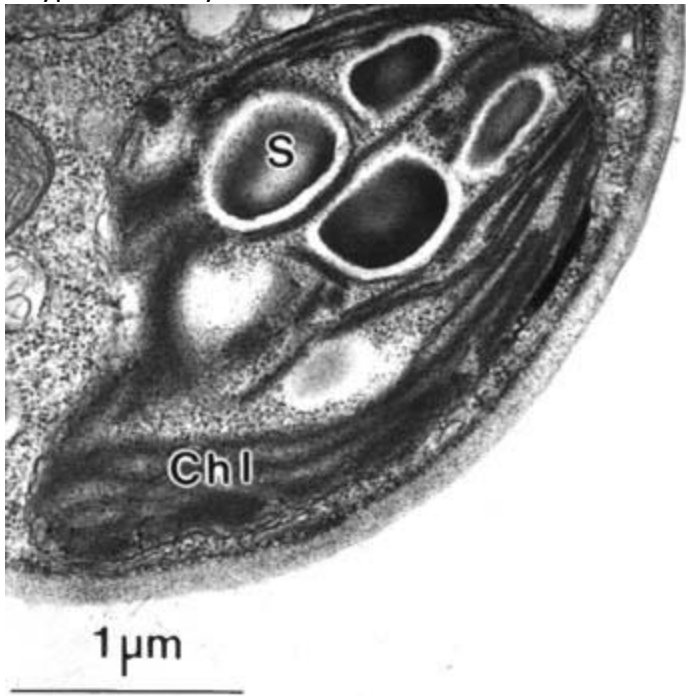
Figure 1



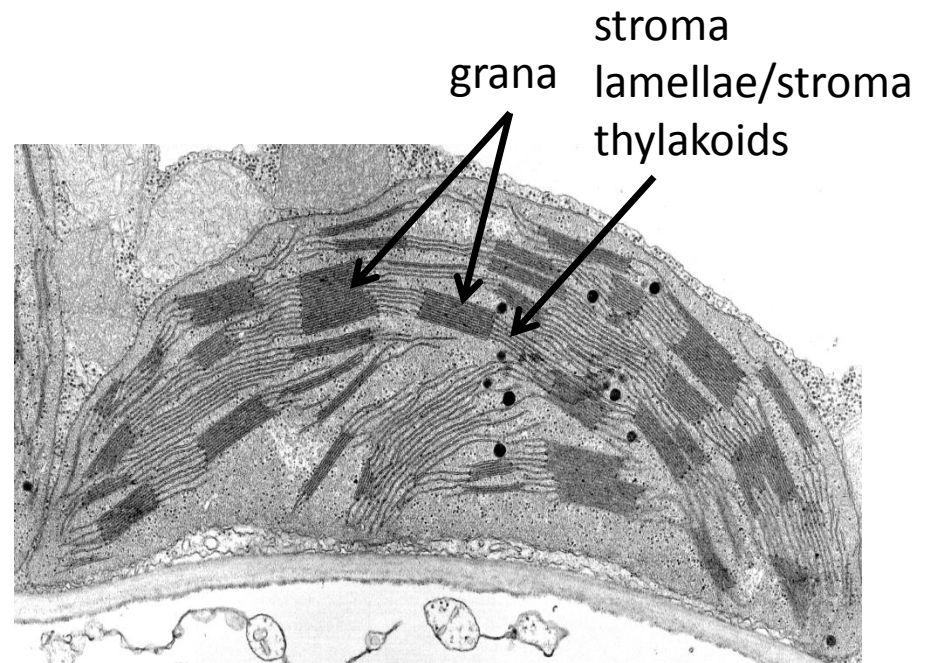
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hypnea.botany.uwc.ac.za

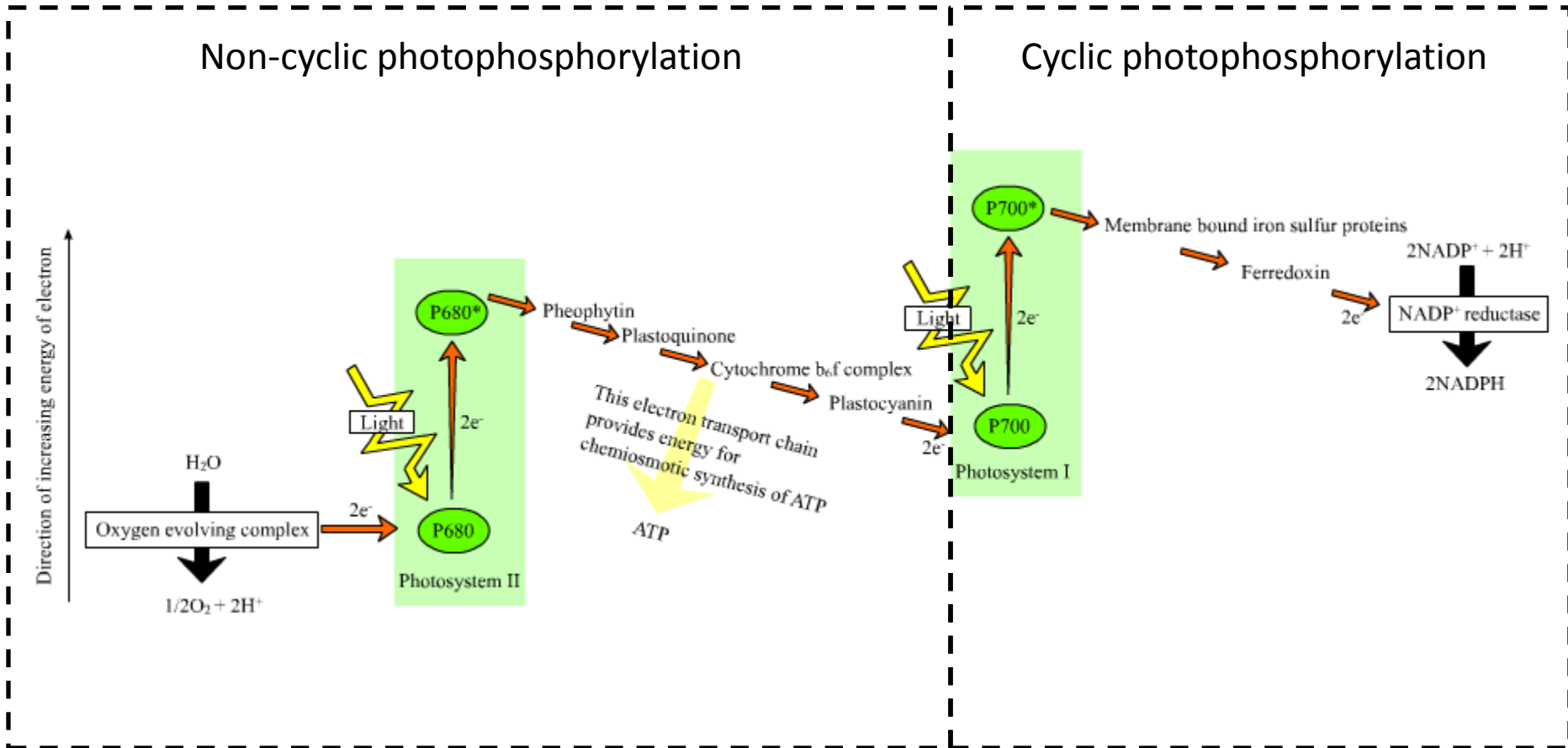


left: Chloroplast of *Chlamydomonas*, a unicellular green alga. Note the chloroplast (Chl) with stacked thylakoids, and starch (s). TEM image by Richard Pienaar

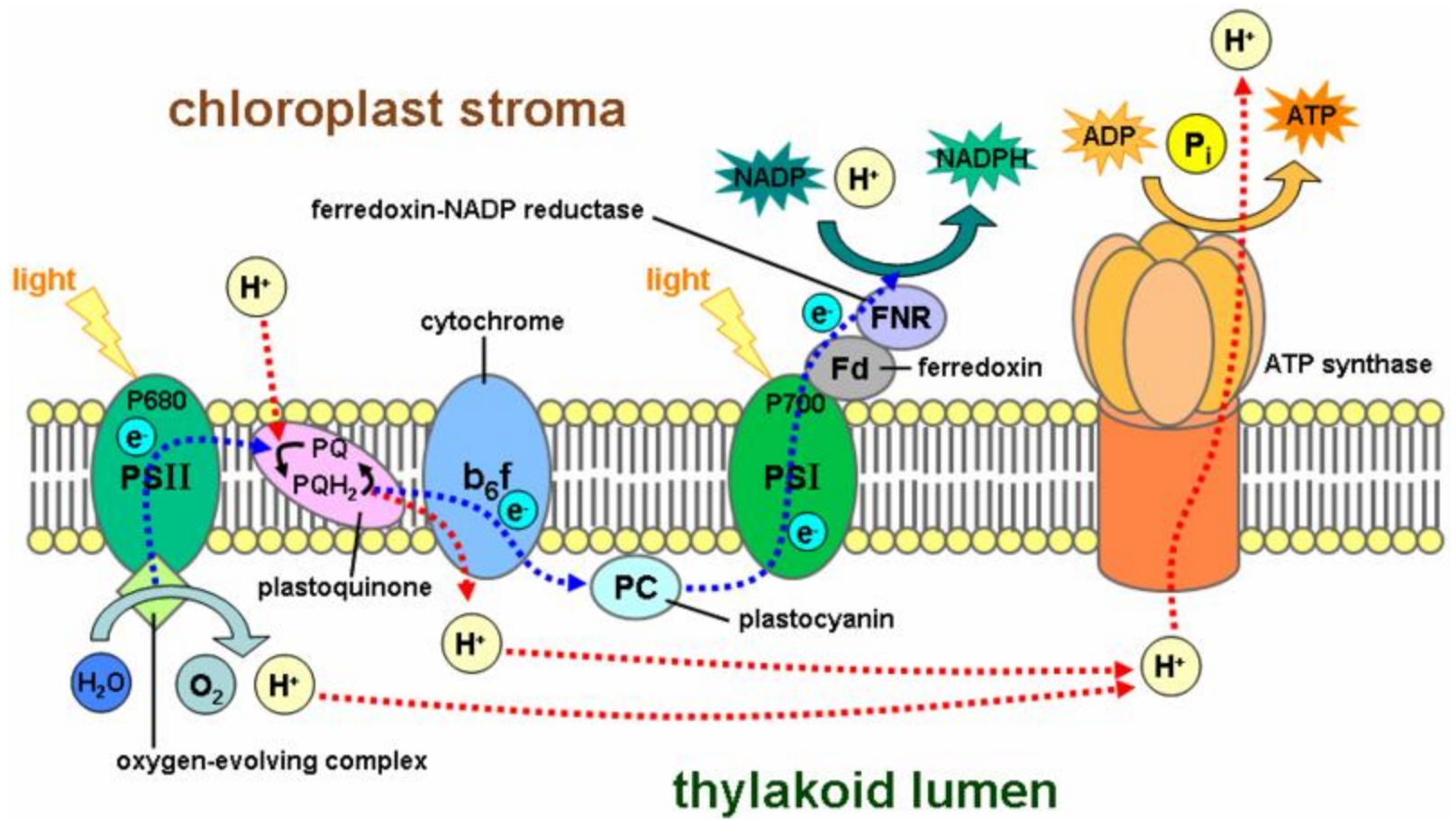


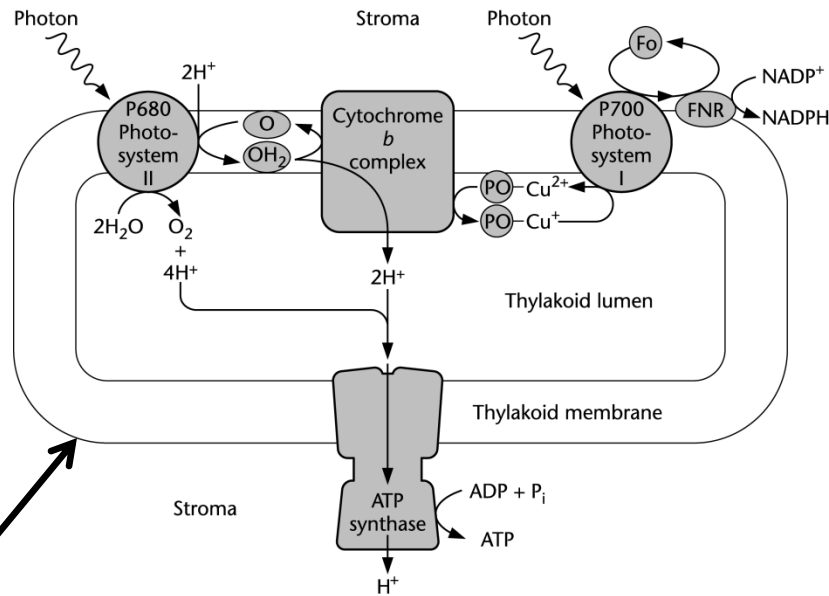


# The 'Z' scheme



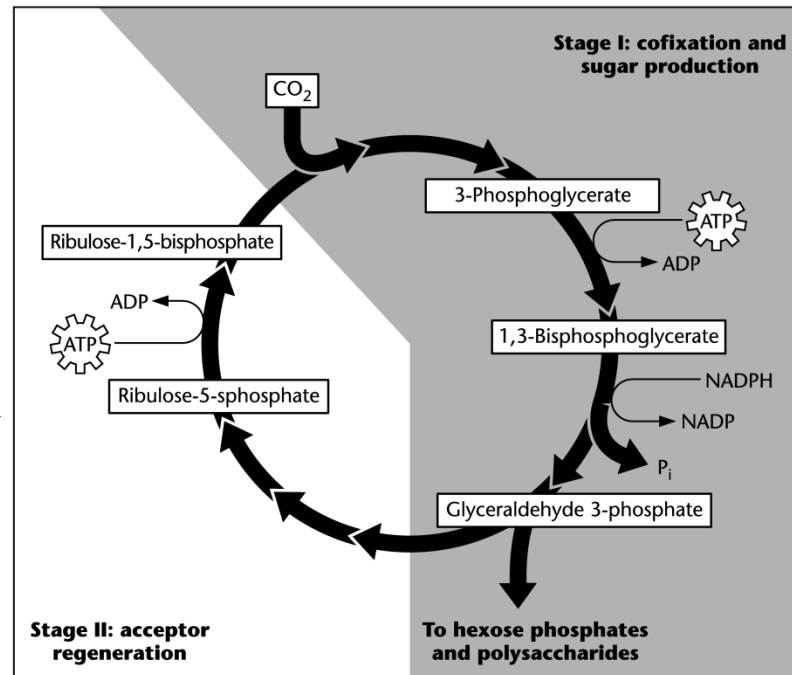
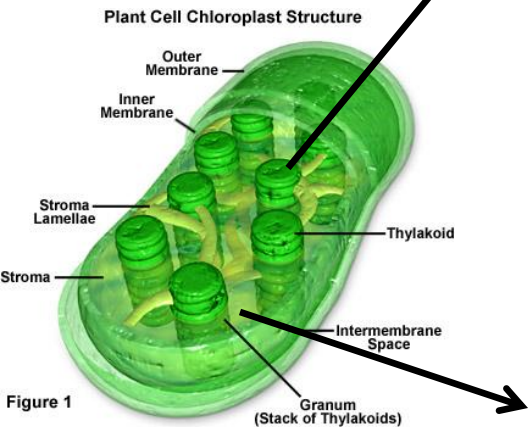
Light reactions:  $O_2$  evolution  
Occurs in thylakoid membranes





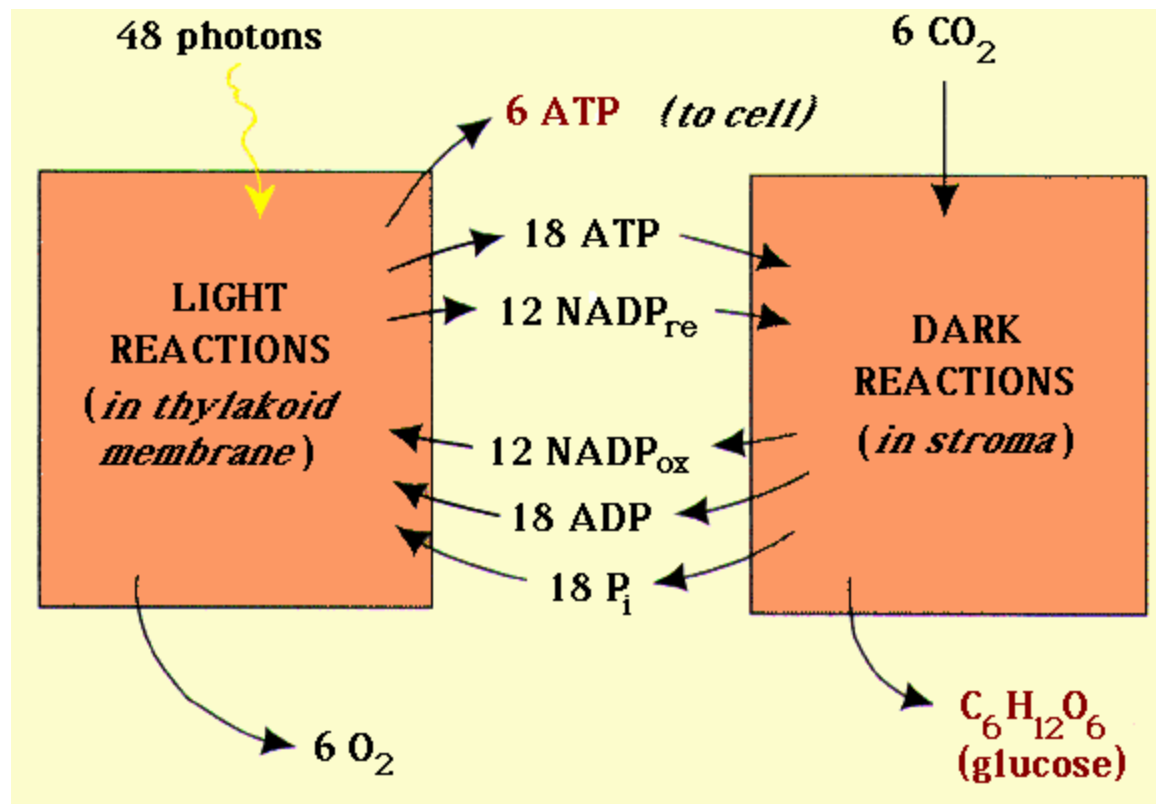
**Light reactions**

*Occurs in thylakoid membrane;  $\text{O}_2$  evolution*

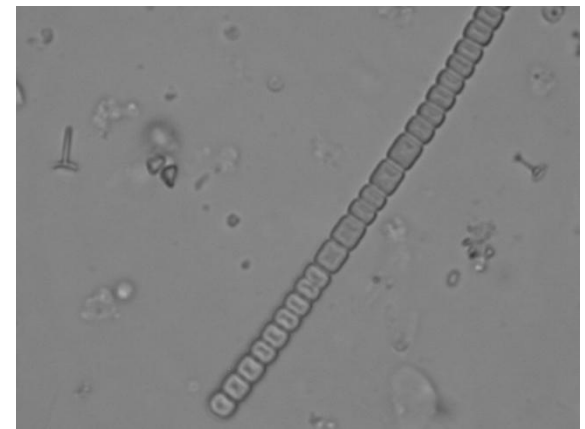
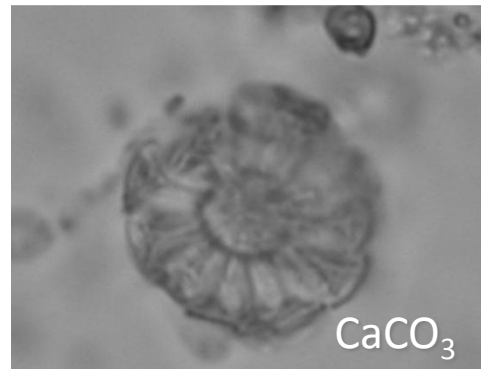
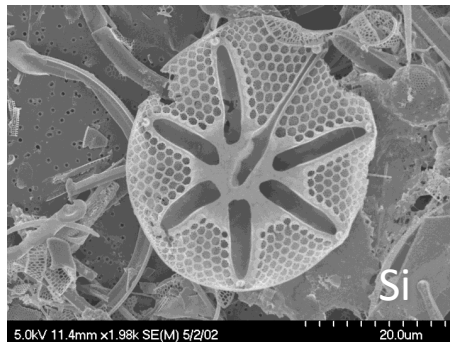
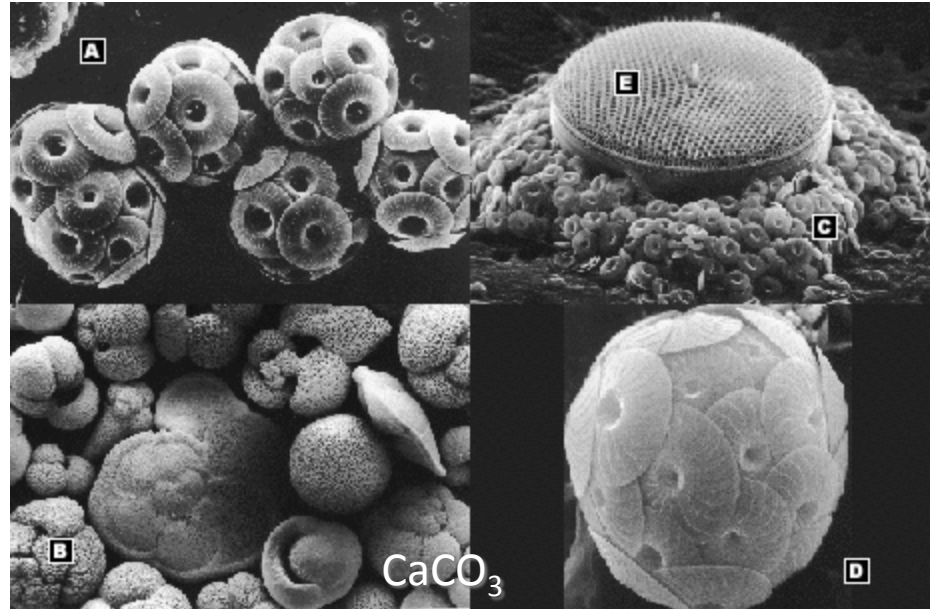
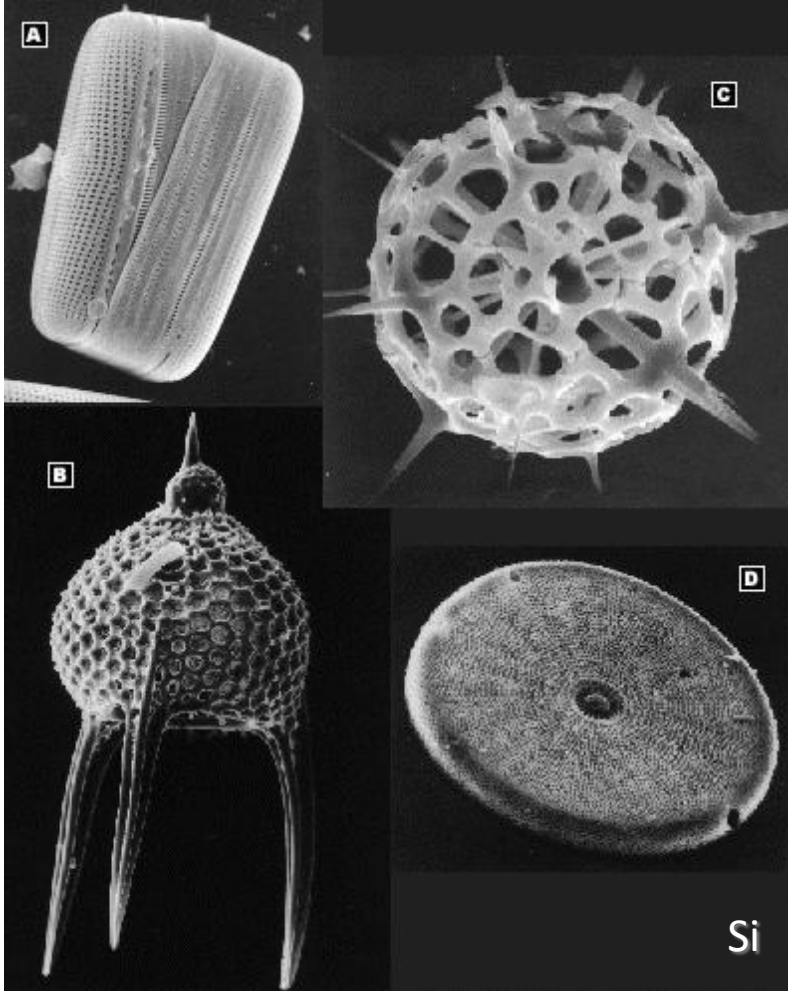


**Dark reactions**

*Occurs in stroma; production of glucose*



# Planktonic forms



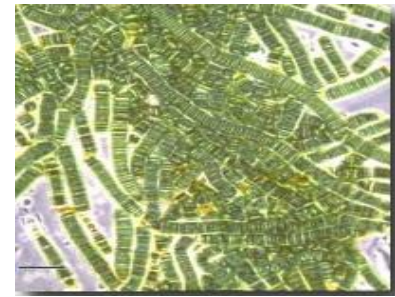
# Classifying phytoplankton

- Size (last time)
- Pigments (photosynthetic pigments, includes chl a, antenna pigments and protective pigments)
- Evolutionary relationships
  - How are they related?
- Functional groups
  - What do they do?

How they are classified depends on how they are measured, and on the question at hand



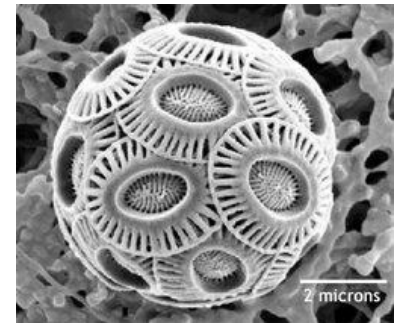
Cyanobacteria oxygenated Earth (>2 billion years ago); harmful algal blooms (more later)



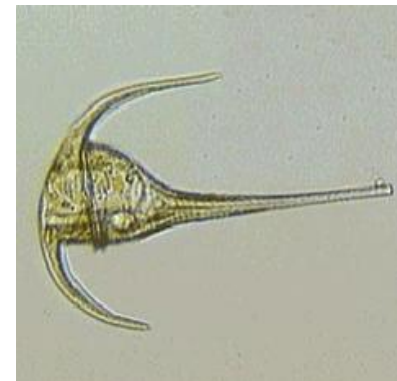
Diatoms great at drawing down CO<sub>2</sub>:  
reduce global temperatures



Coccolithophores alter and are altered  
by ocean's pH; produce DMS (trace gas  
connected to climate)



Dinoflagellates – harmful algal  
blooms, extraordinary diversity of  
form & function



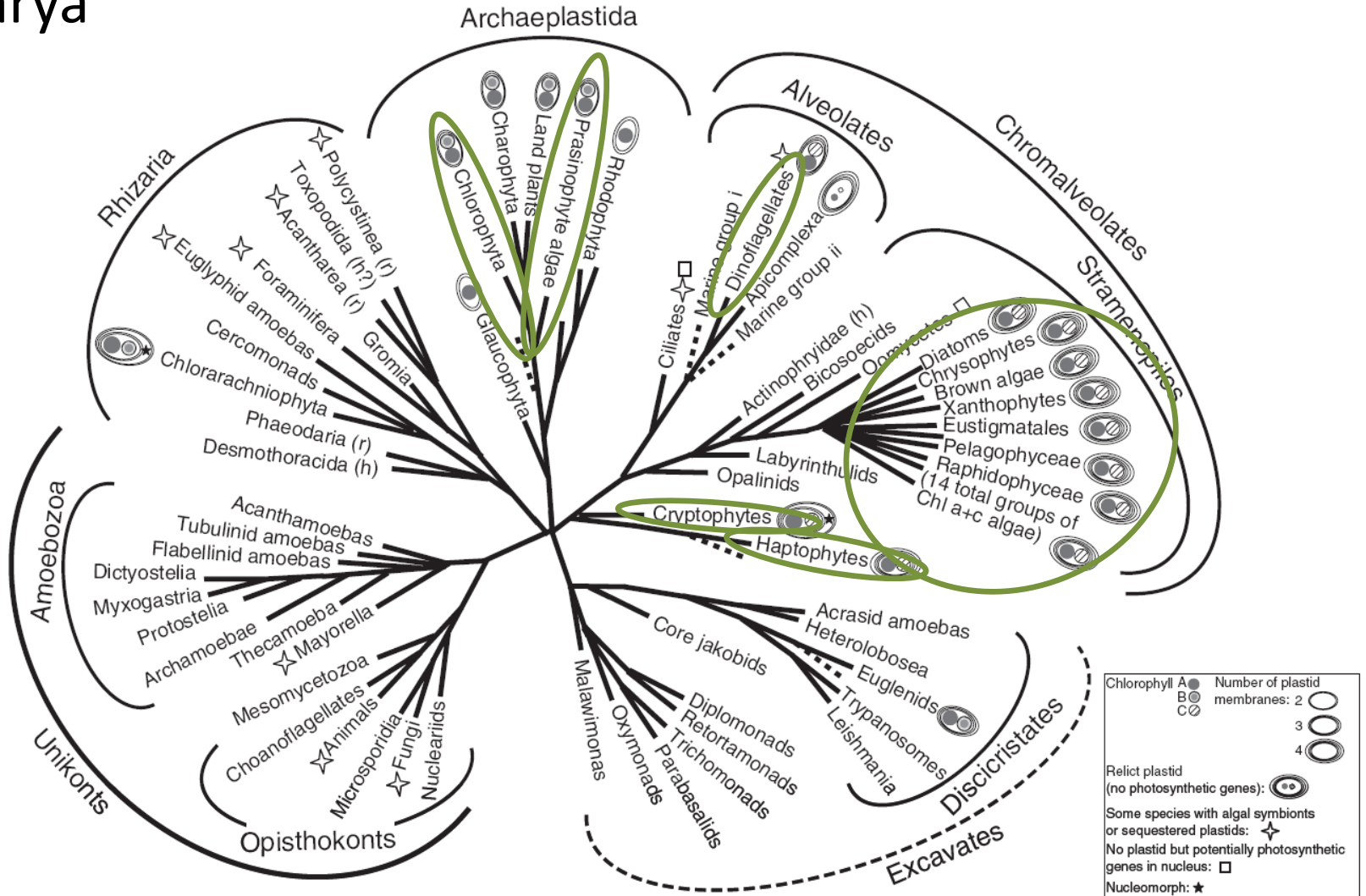
# Phytoplankton in the eukaryotic tree of life

- Phylogeny/taxonomy is complicated and continuously being revised; see Adl et al. 2005 (on website) for recent classification scheme

## Domain Eukarya

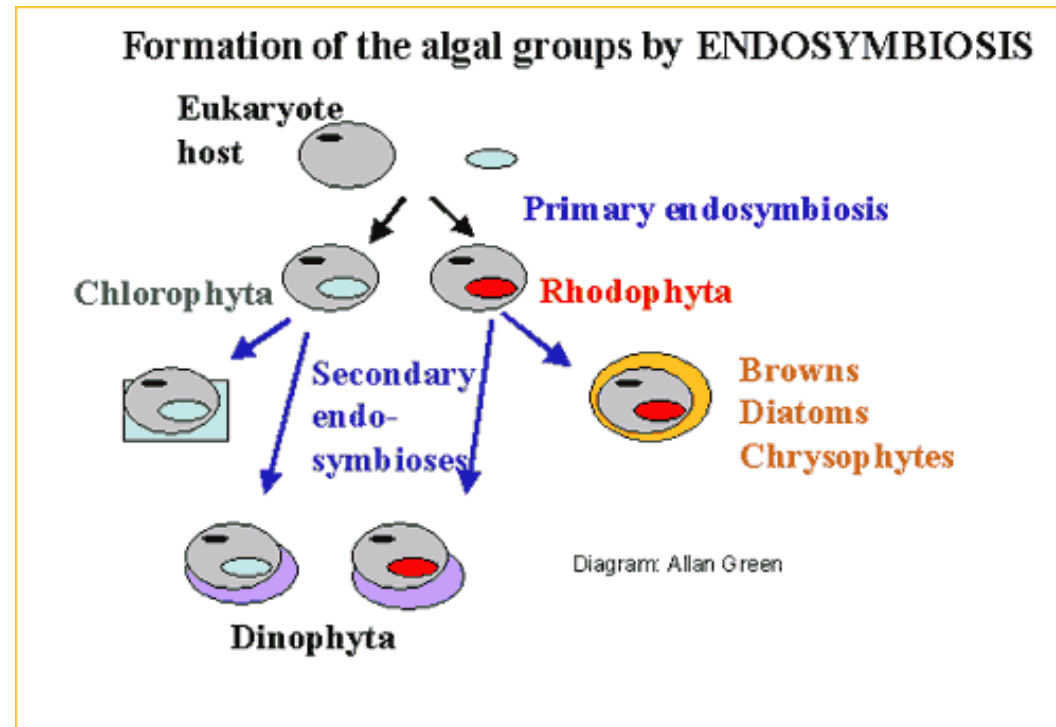
Supergroup	First rank	Second rank (examples)
Archaeplastida	Chloroplastida	Chlorophyta, Prasinophyta
Chromalveolata	Cryptophyceae	Cryptomonadales
	Haptophyta	Pavlovophyceae, Prymnesiophyceae
	Stramenopiles (= Heterokontophyta)	Bacillariophyta, Chrysophyceae, Dictyochophyceae, Pelagophyceae, Raphidophyceae, Synurales, Xanthophyceae, Dinzoa
Excavata	Euglenozoa	Euglenida

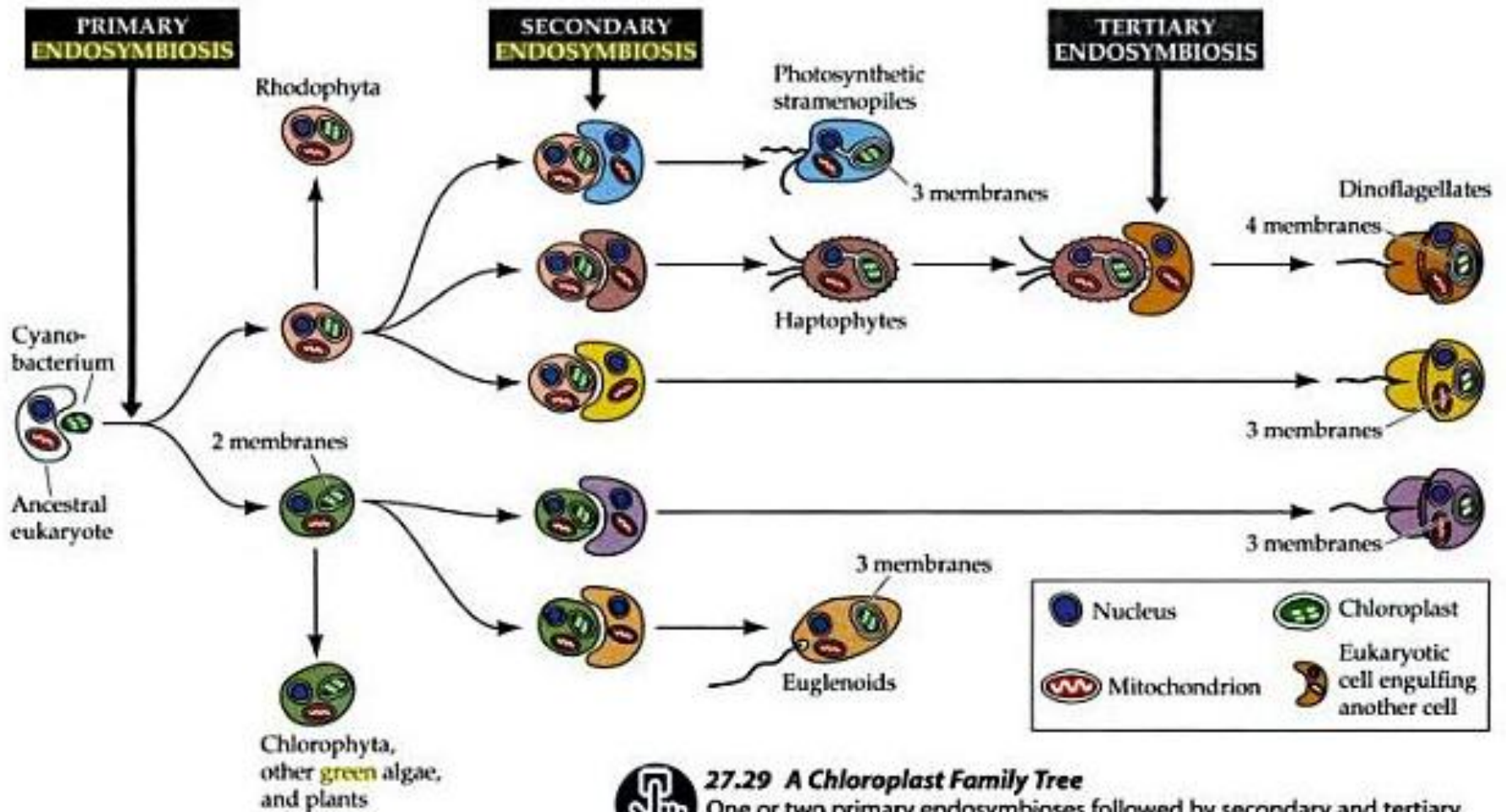
# Eukarya



# Evolutionary relationships

- Endosymbiotic theory (serial endosymbiosis)
- Eukaryote engulfed a prokaryote
  - Photosynthetic: chloroplast;
  - nonphotosynthetic: mitochondrion
  - Green and red lineages





**27.29 A Chloroplast Family Tree**

One or two primary endosymbioses followed by secondary and tertiary endosymbioses gave rise to all of today's chloroplasts.

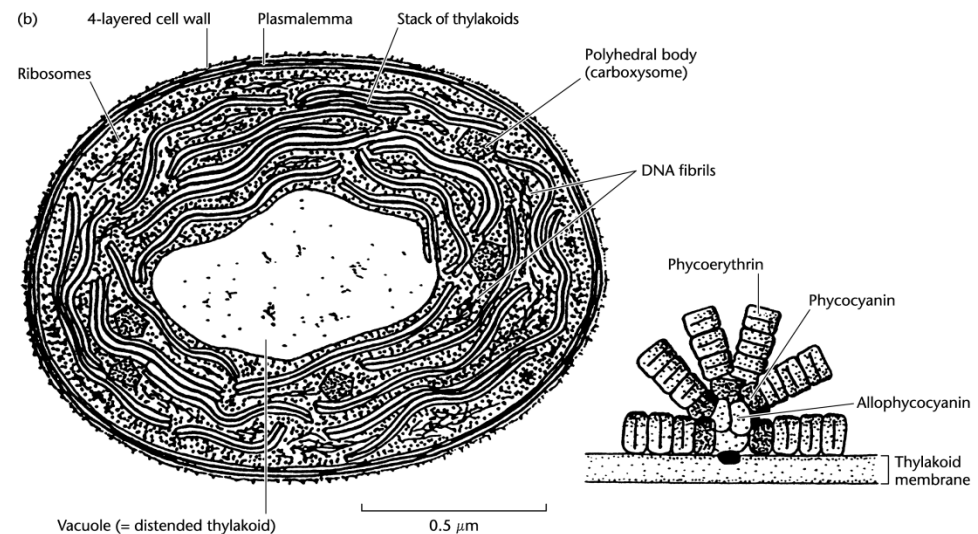
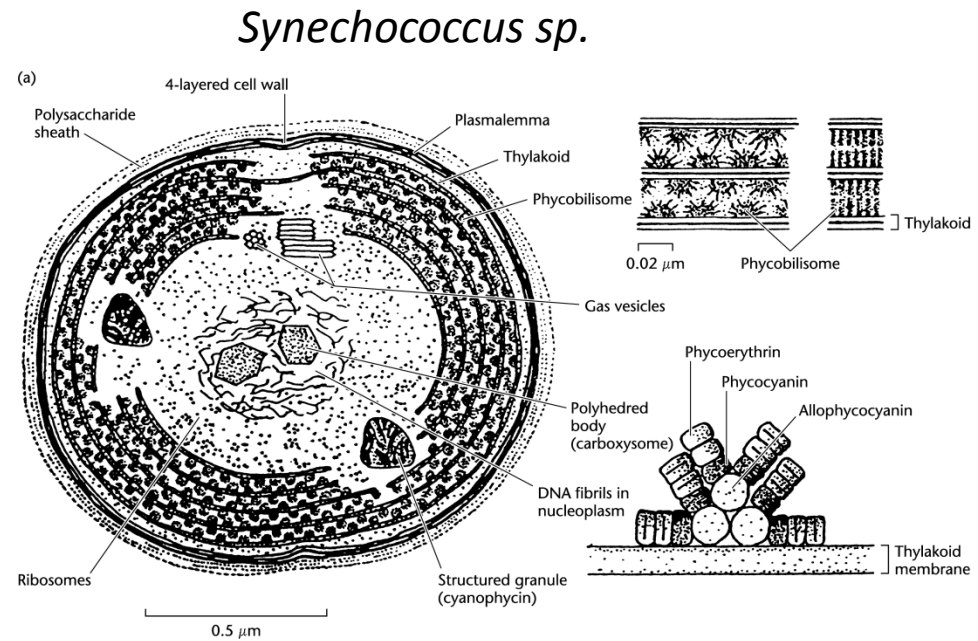
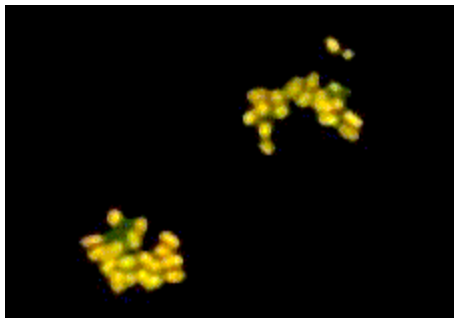
# Major classes of phytoplankton

- Bacteria
  - Cyanophyceae and Prochlorophyceae (Bacteria)
- Eukarya
  - **Archaeplastida**: Chloroplastida (Chlorophyceae, Prasinophyceae)
  - **Chromalveolata**: Cryptophyceae (Cryptomonadales), Haptophytes (Pavlophyceae, Prymnesiophyceae), Heterokonts (Bacillariophyta, Chrysophyceae, Dictyochophyceae, Pelagophyceae, Raphidophyceae, Synurales, Xanthophyceae, Dinzoa)
  - **Excavata**: Euglenozoa (Euglenida/Euglenophyceae)



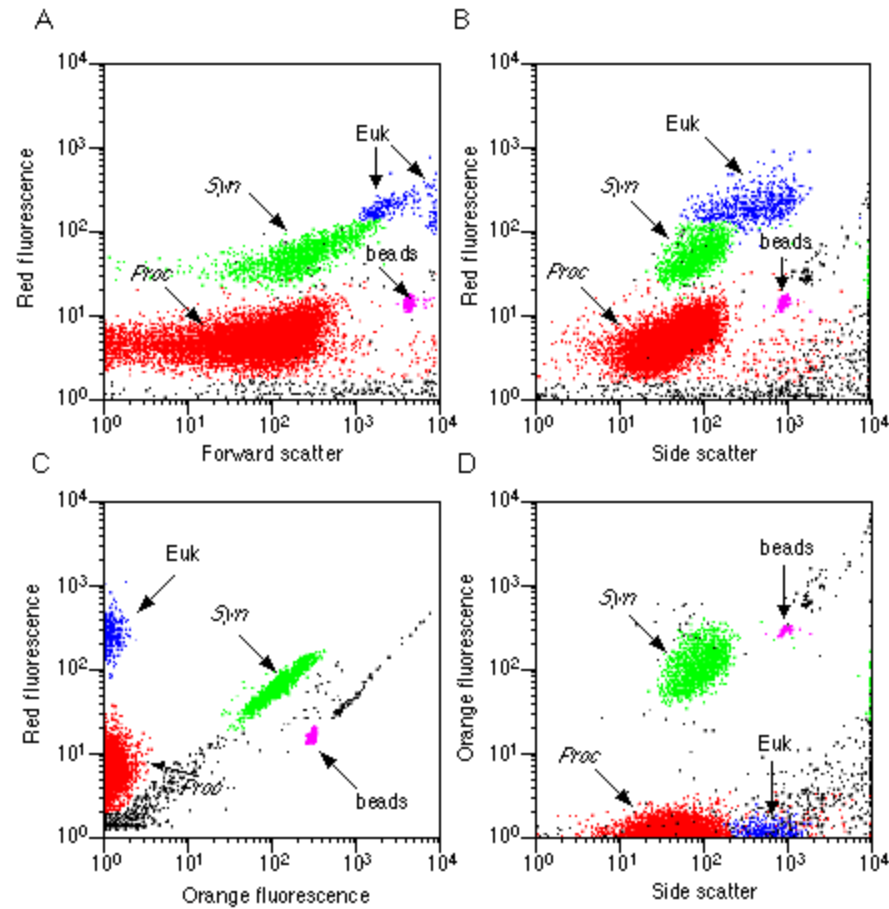
# Cyanobacteria

- Bacteria
- Chlorophyll, but no chloroplasts
- Syn: Phycocyanin, Phycoerythrin, allophycocyanin
- Pro: divinyl chlorophyll b
- Cell wall of murein

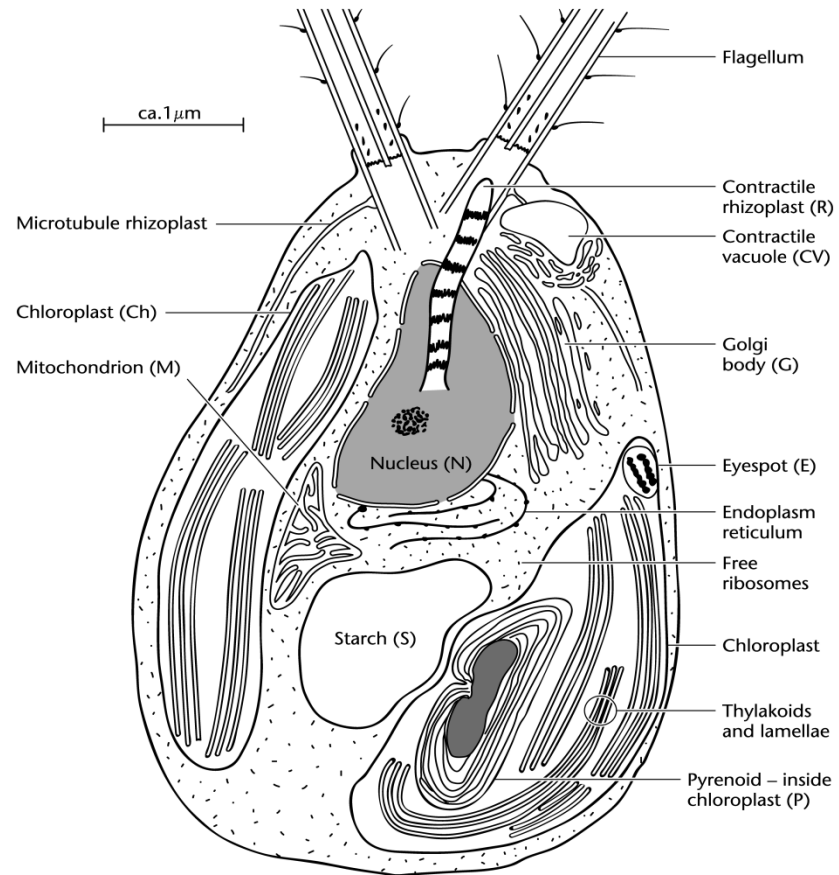


*Prochlorococcus marinus*

# *Synechococcus* and *Prochlorococcus* as viewed by a flow cytometer

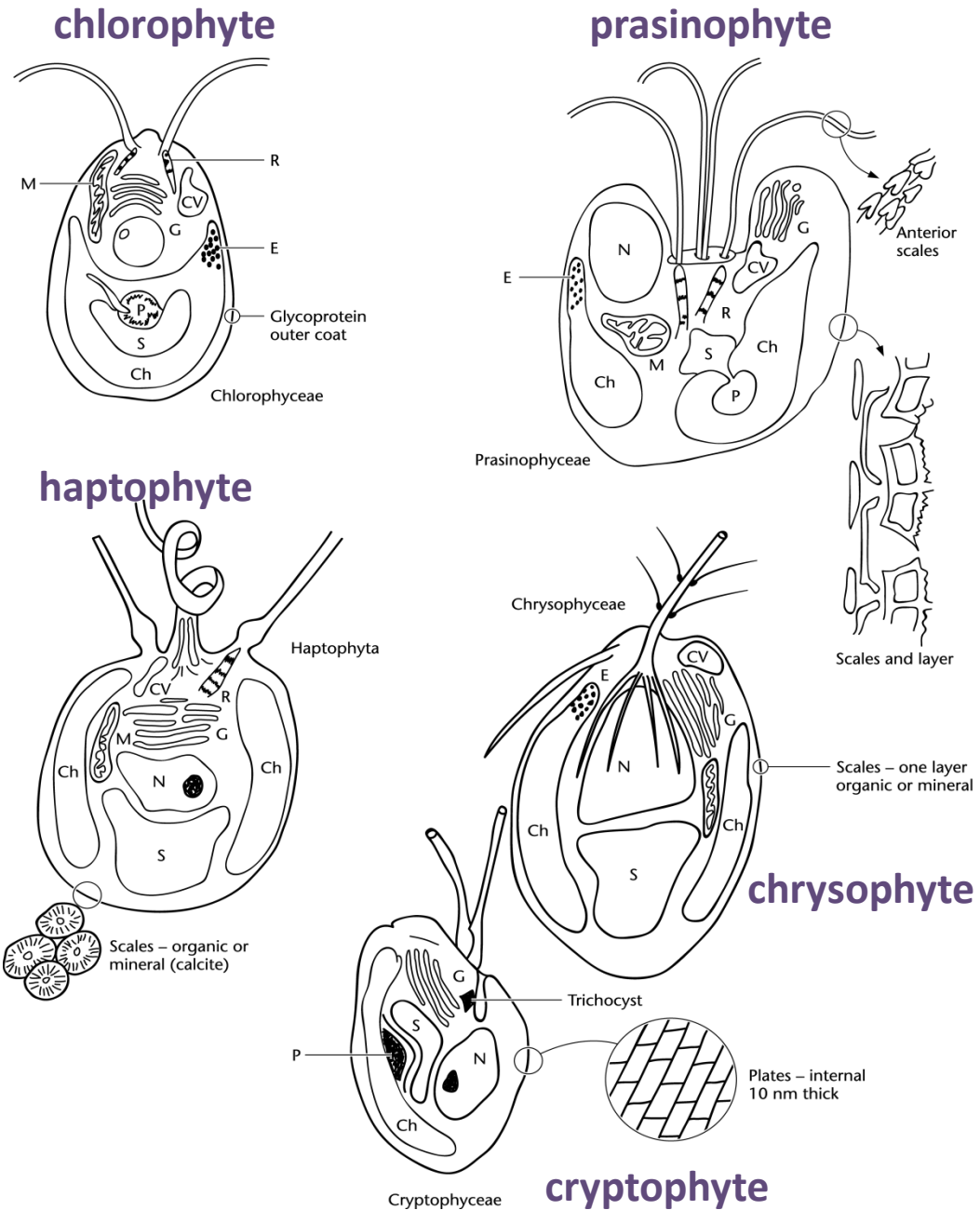


# Generalized “microflagellate”



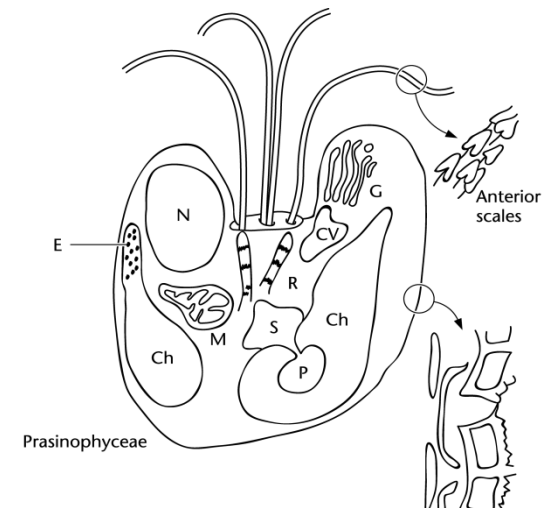
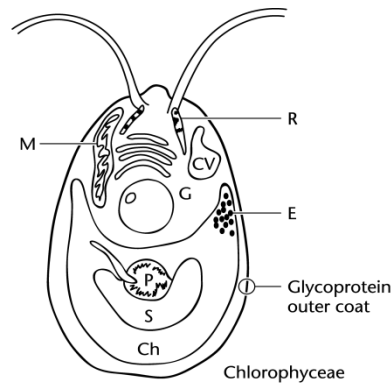
## Types of microflagellates

- Differ according to pigments, cell wall material, number of flagella
- Very diverse group
- Artificial grouping (not related – polyphyletic, many origins)
- If classified based on size, they have similar characteristics; if measured by pigments, can be quite different



## Archaeplastida: Chloroplastida (Chlorophyceae, Prasinophyceae)

- possess chl *a* and chl *b*
- possess flagella that are similar in structure, can have two or more
- chloroplast is surrounded by two membranes
- accessory pigments include lutein, zeaxanthin, violaxanthin, anteraxanthin, neoxanthin
- some possess pyrenoids (proteinaceous product involved in starch formation)
- storage product is starch, present as grains



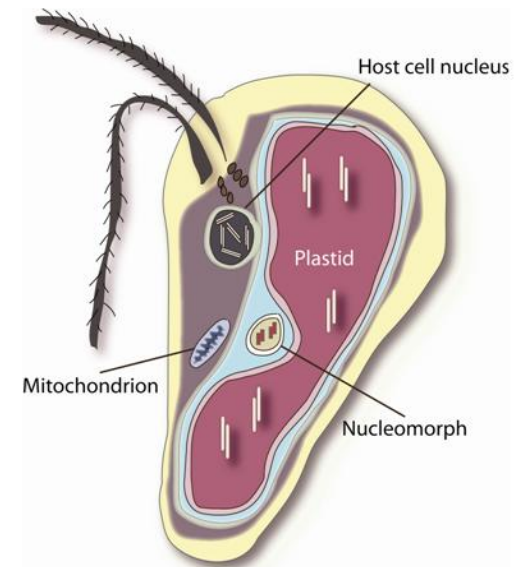
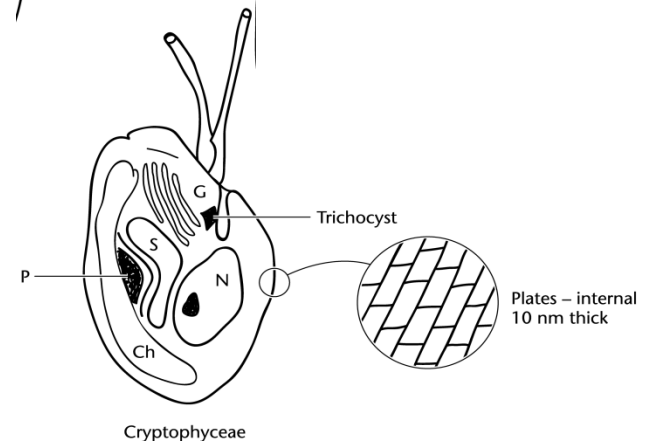
# Chromalveolata

- **Cryptophyceae** (Cryptomonadales)
- Haptophytes (Pavlophyceae, **Prymnesiophyceae**)
- Heterokonts (**Bacillariophyta**, Chrysophyceae, Dictyochophyceae, Pelagophyceae, Raphidophyceae, Synurales, Xanthophyceae, **Dinzoa**)

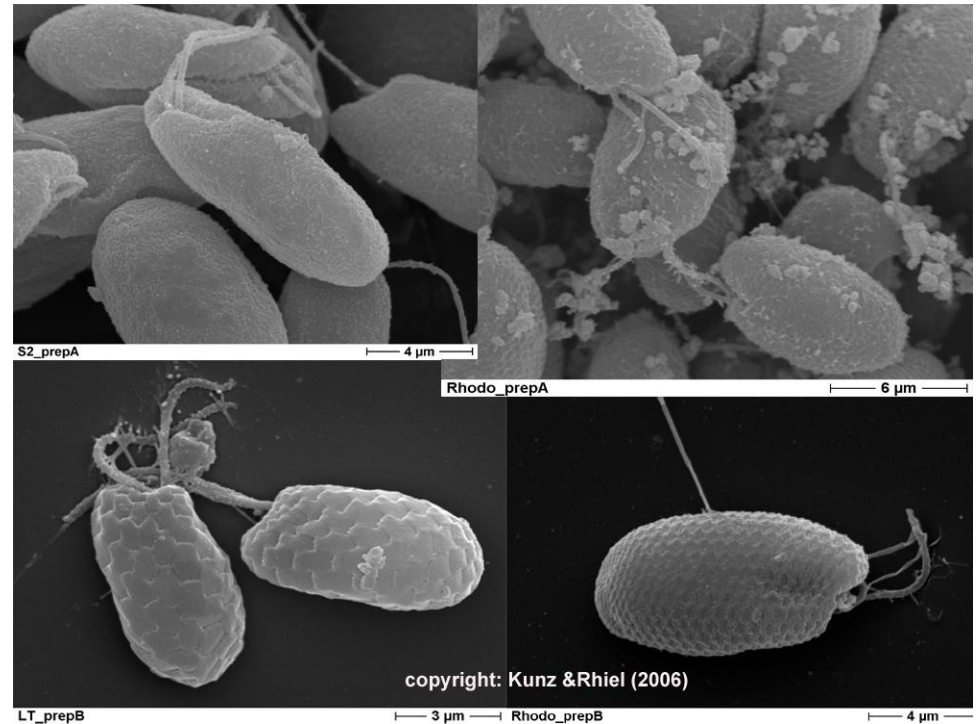


- Two flagella which differ in length
  - longer flagellum bears two rows of mastigonemes (stiff lateral hairs), shorter flagellum bears a single row of shorter mastigonemes
  - flagella covered with tiny organic scales
- Dorsal side is convex, while ventral side is flat, with a shallow longitudinal groove
- Gullet on the ventral side at the anterior end of the flagellar groove – gullet is lined with trichocysts (releases small protein thread into medium for rapid movement)
- Chloroplasts surrounded by a fold of endoplasmic reticulum
- Chloroplasts contain chl  $a$  and  $c_2$ ; accessory pigments include phycocyanin, phycoerythrin,  $\alpha$ -carotene, alloxanthin, zeaxanthin, monadoxanthin, crocoxanthin
- No phycobilisomes

# Cryptophycee



- Storage material is starch
- Cell is enclosed by stiff, proteinaceous periplast, usually made up of rectangular or polygonal plates
- A nucleomorph is present in the space between the chloroplast and the chloroplast ER (contains DNA and a nucleolus-like structure; interpreted as the vestigial nucleus of a photosynthetic eukaryotic endosymbiont)
- Found in both fresh and marine waters

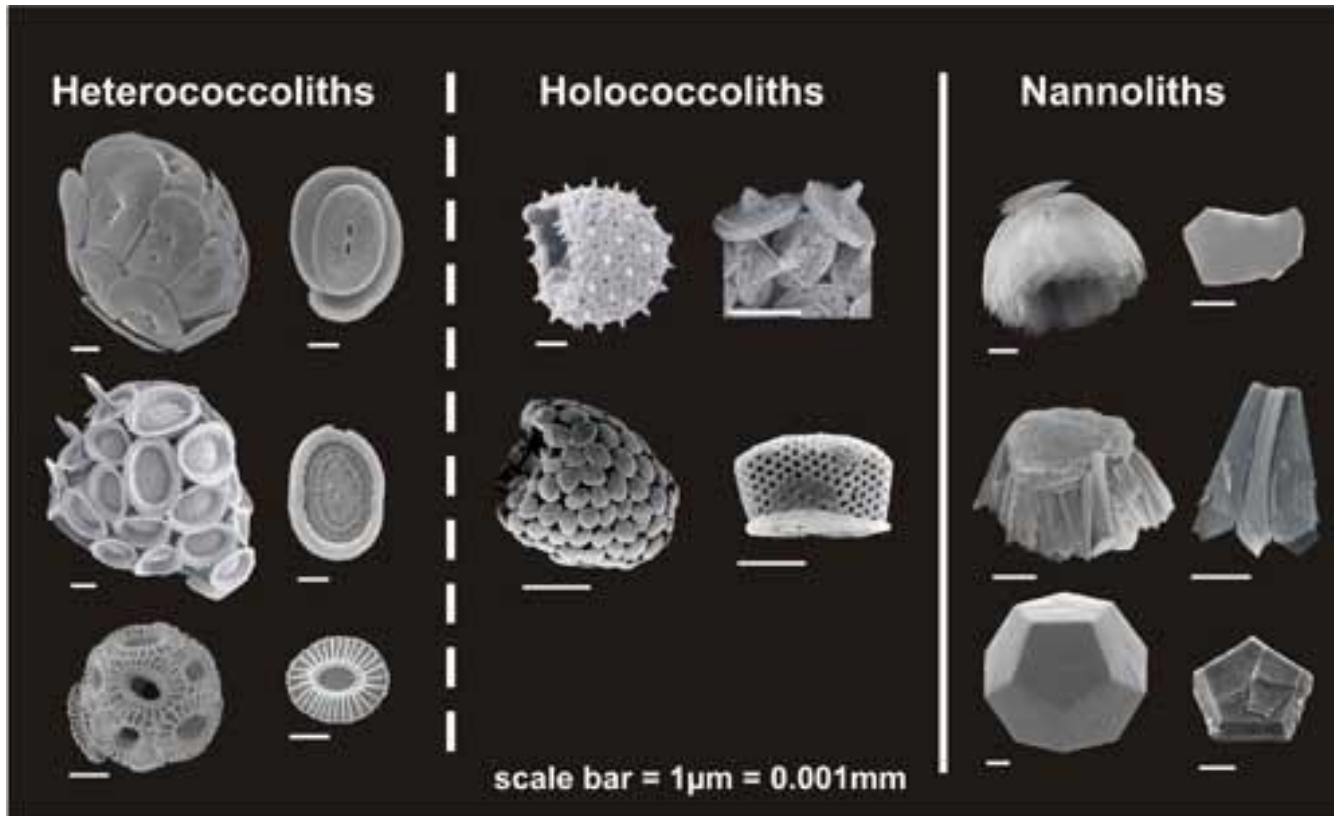


## Cryptophyceae

# Haptophytes (= Prymnesiophytes)

- Majority are unicellular
- Possess two flagella without mastigonemes plus a haptonema (thin, filamentous appendage which may be short or very long – not a flagellum, different structure)
- Often possess pyrenoids
- Contain chl a, c1 and/or c3, c2; accessory pigments include 19'hexanoyloxyfucoxanthin, 19'butanoyloxyfucoxanthin, and fucoxanthin, plus b-carotene, diadinoxanthin, and diaxanthin
- Storage product is chrysolaminarin (polysaccharide)
- Cell surface covered in scales, granules of organic matter (cellulose), or else calcified scales (coccoliths)

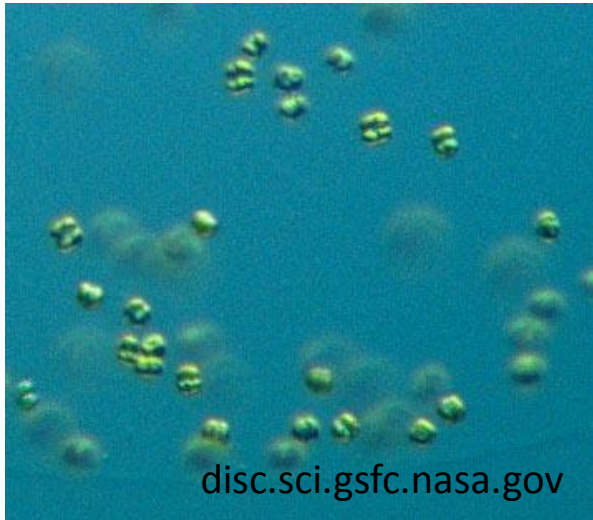
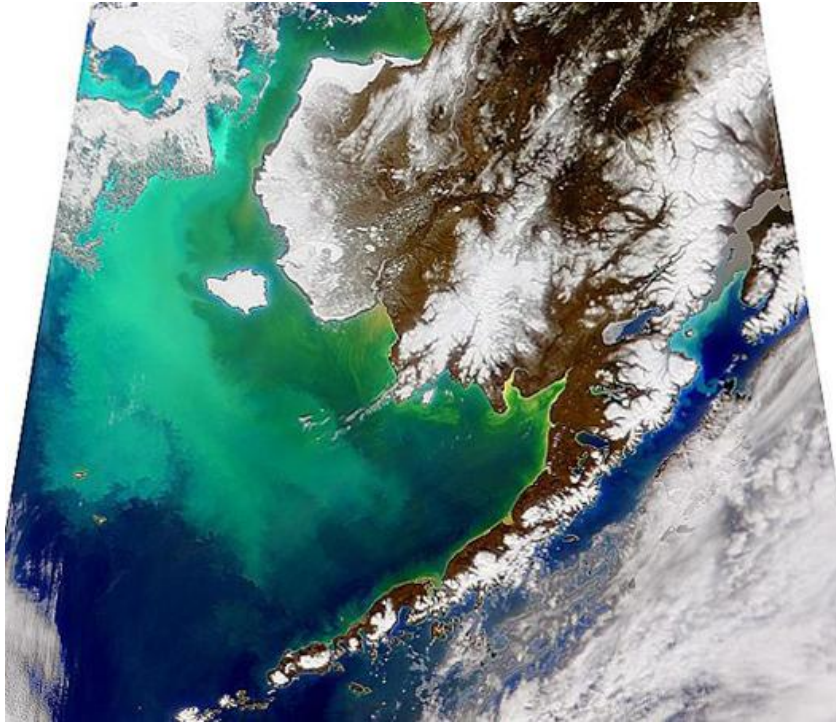
# Haptophytes



*Chrysochromulina* sp.  
[www.glerl.noaa.gov](http://www.glerl.noaa.gov)



# Haptophytes

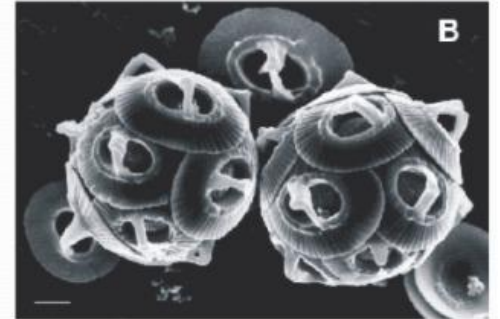
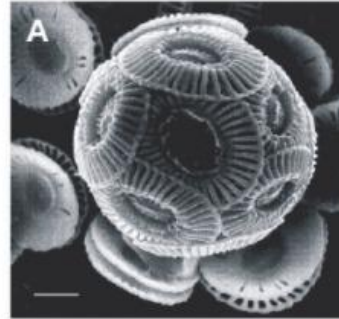


[disc.sci.gsfc.nasa.gov](http://disc.sci.gsfc.nasa.gov)

*Emilianaia huxleyi*

*Gephyrocapsa oceanica*

300 ppmv CO<sub>2</sub>



800 ppmv CO<sub>2</sub>



*Riebesell et al.*

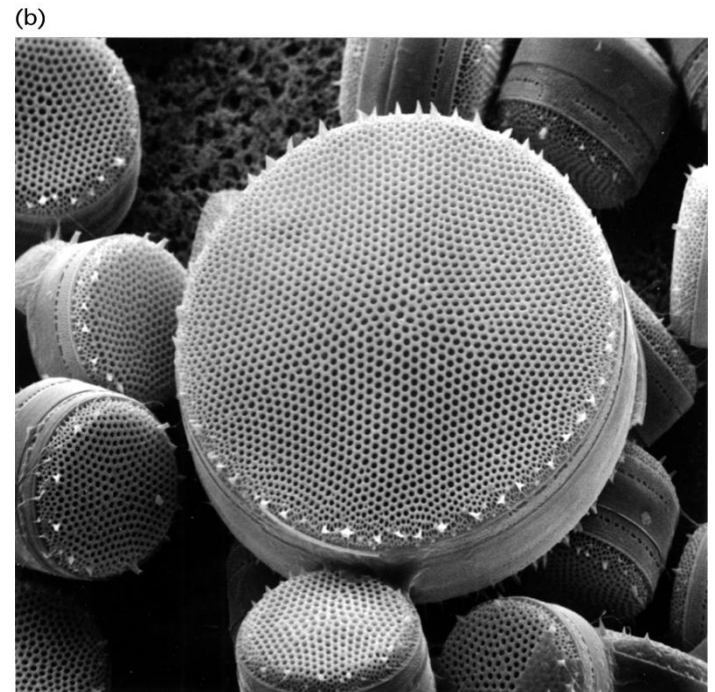
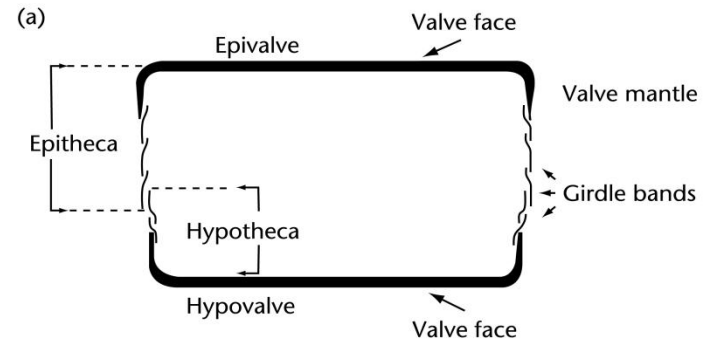
# Heterokontophyta/Stramenopiles

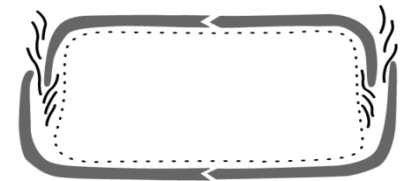
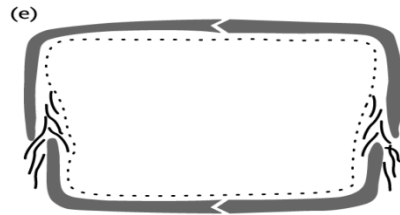
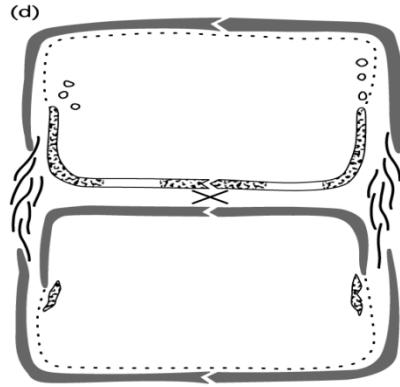
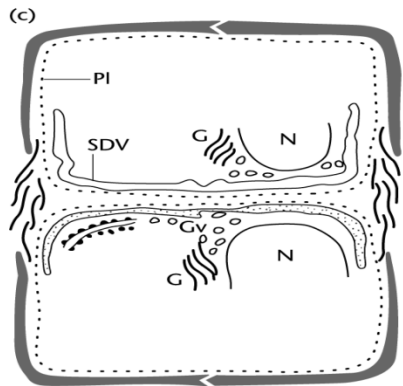
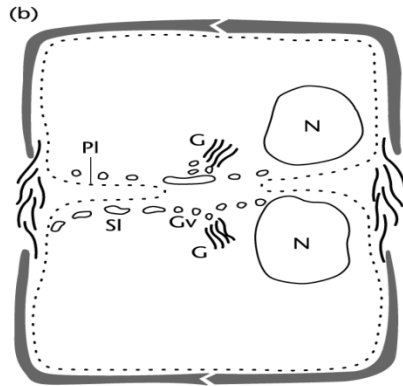
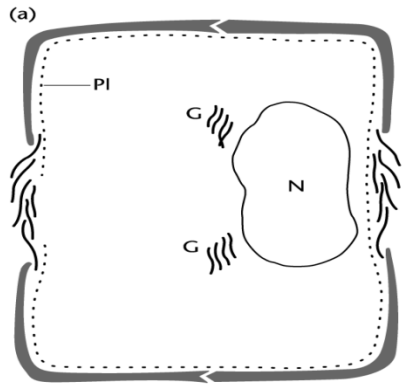
- Heterokonts have two flagella, one long pleuronematic (= flimmer or tinsel) flagellum with stiff hairs (mastigonemes) directed forward during swimming and a shorter smooth flagellum that points backwards along the cell
- Chloroplast enclosed by double membrane and by a fold of endoplasmic reticulum (called chloroplast ER)
- Chloroplasts contain chl  $a$ ,  $c_1$  and  $c_2$ , with fucoxanthin as principal accessory pigment in some groups (Chrysophyceae, Bacillariophyceae, some Raphidophyceae) or vaucheriaxanthin in other groups
- Main reserve polysaccharide is chrysolaminarin ( $\beta$ -1,3 linked glucan) found outside the chloroplasts inside special vacuoles

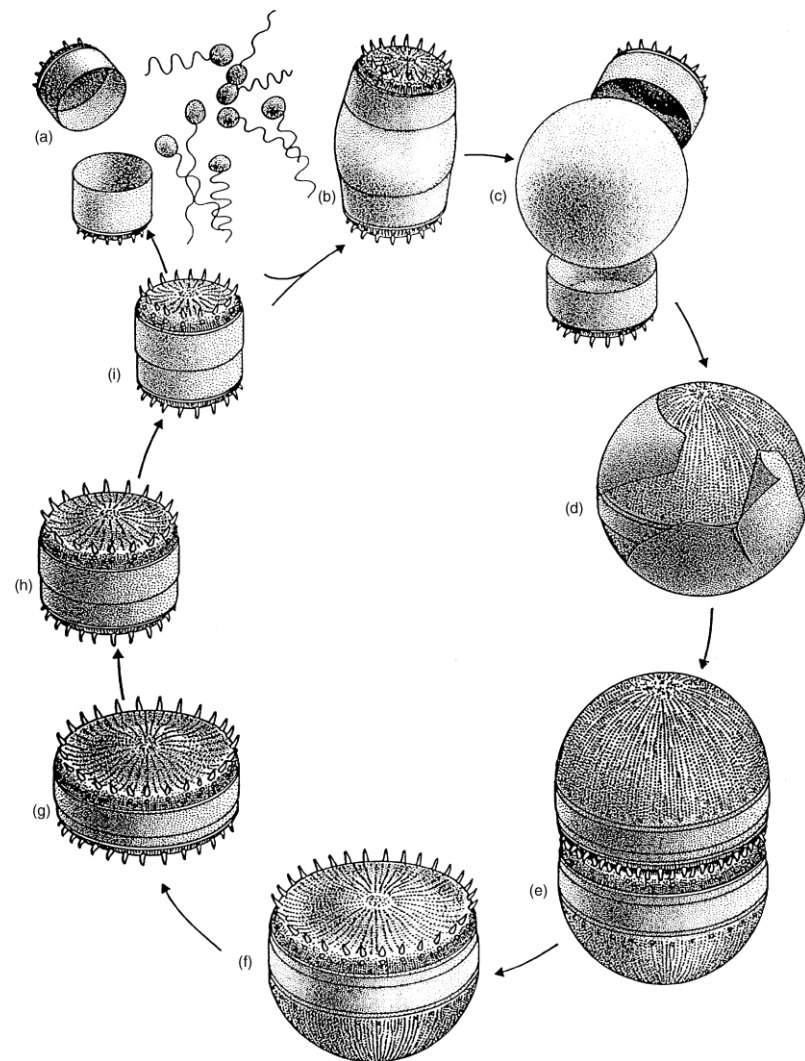
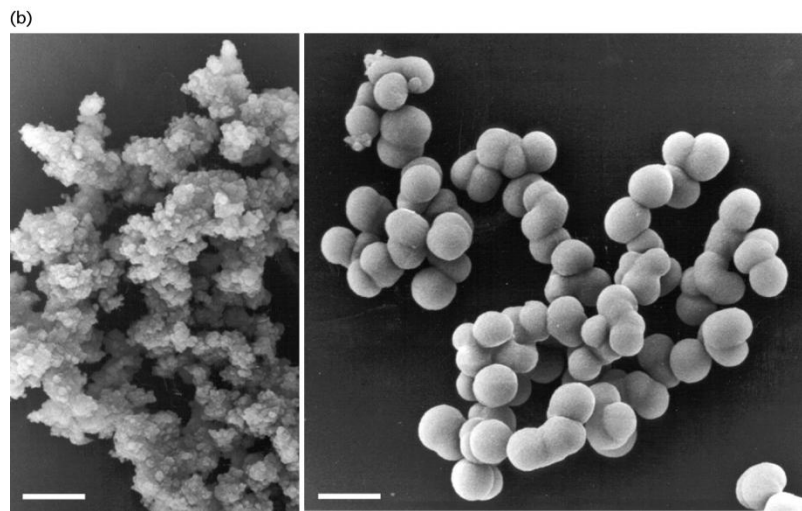
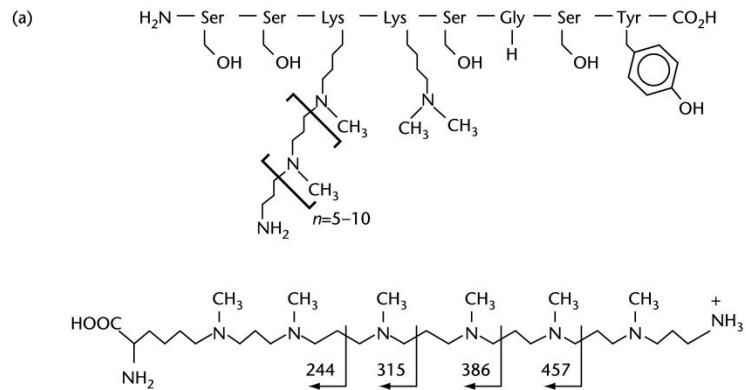


# Diatoms

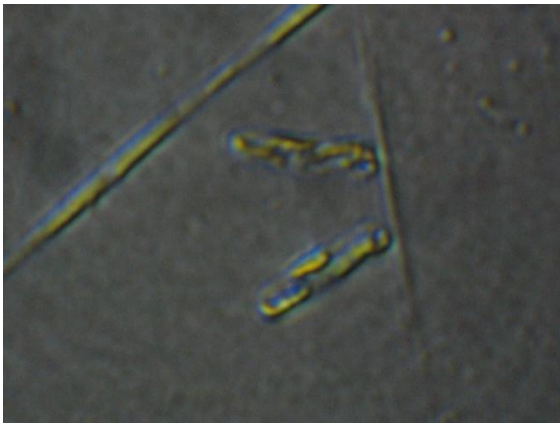
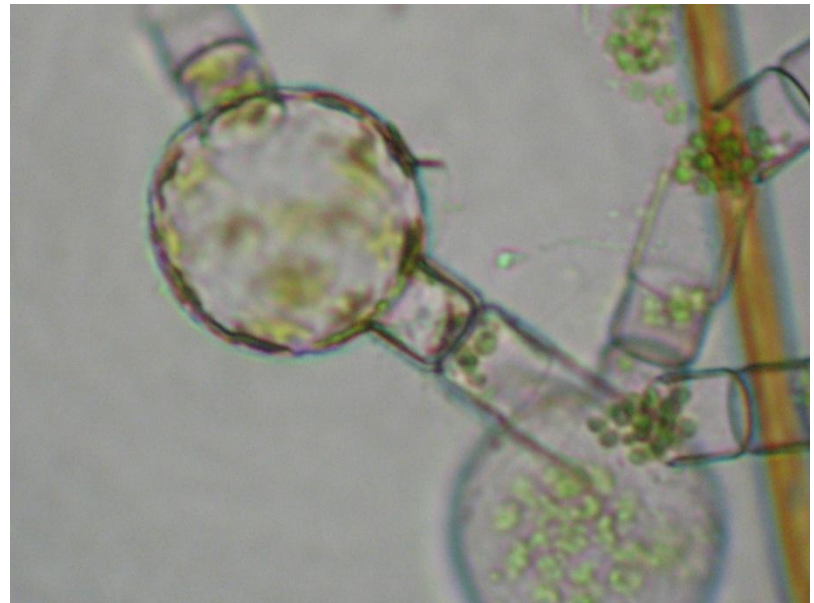
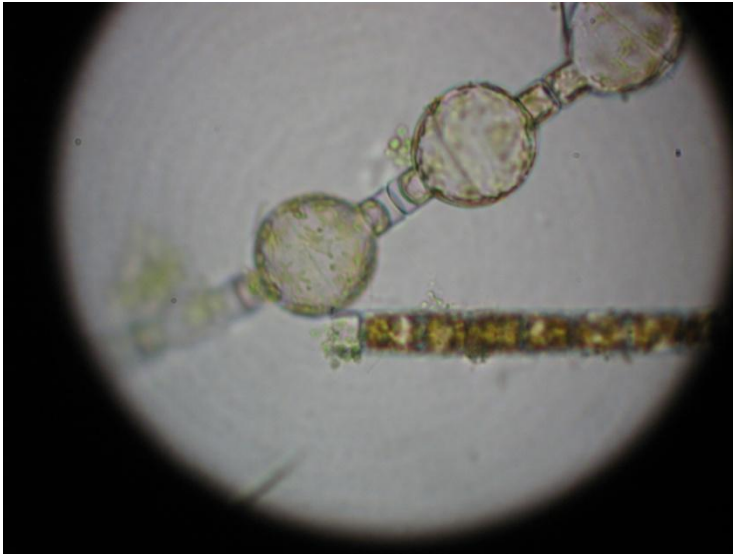
- Bacillariophyceae
- Pigments: chl  $a$ , chl  $c_2$ , fucoxanthin, diadinoxanthin,  $\beta$ -carotene, diatoxanthin, chl  $c_1$
- Cell wall: amorphous silica (opal)
- Centric (radially symmetric) and pennate (bilaterally symmetric)





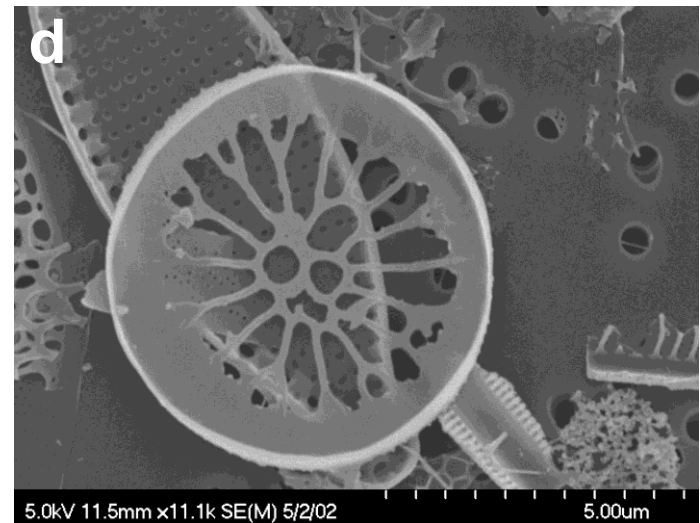
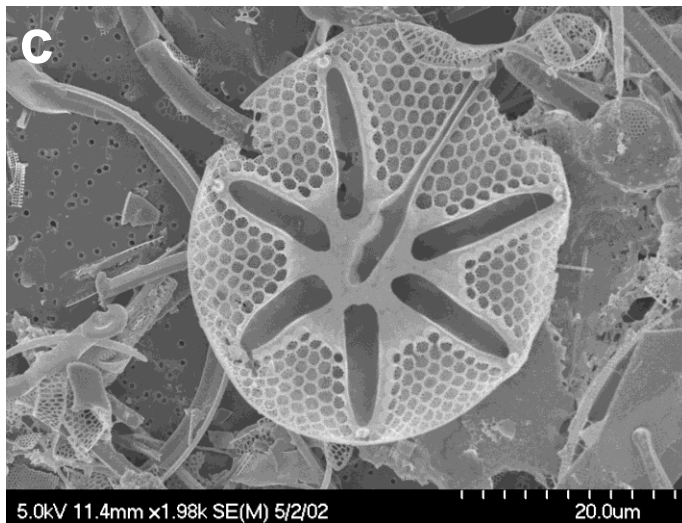
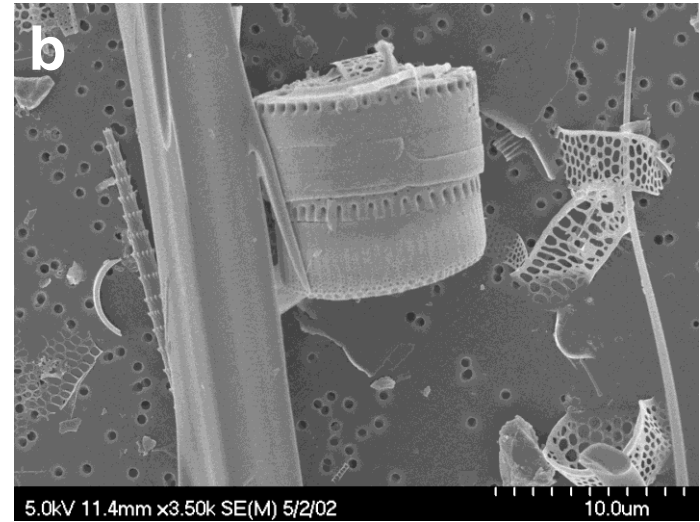
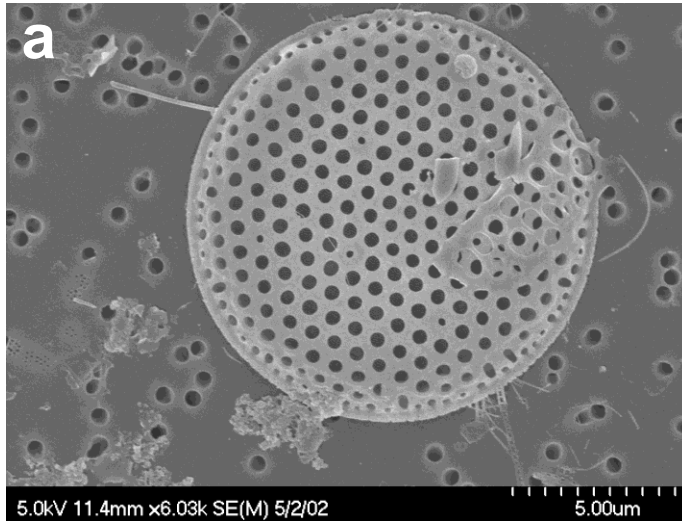


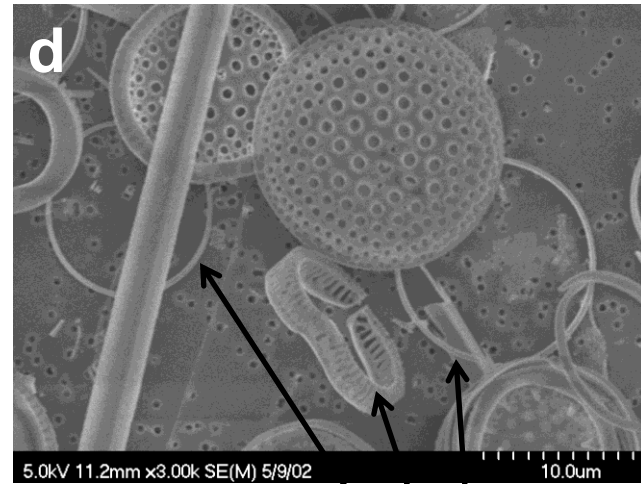
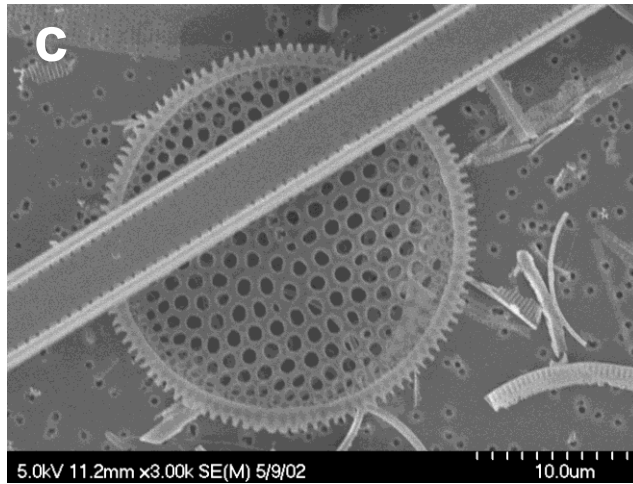
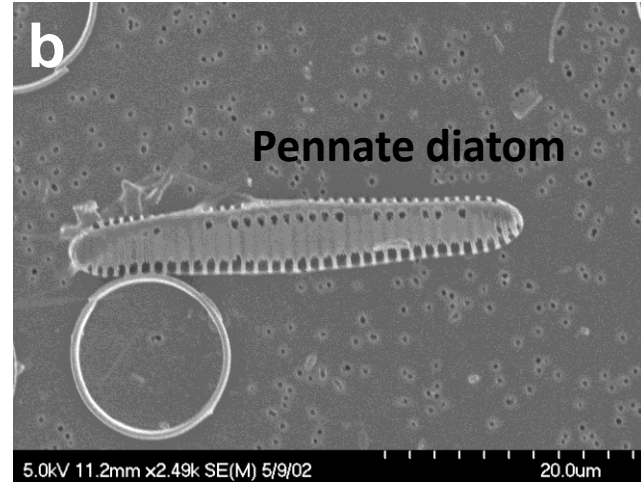
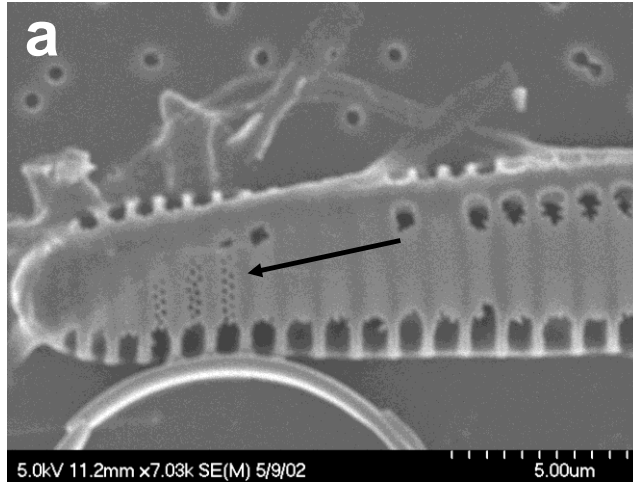
Auxospores of *Aulacoseira* sp.



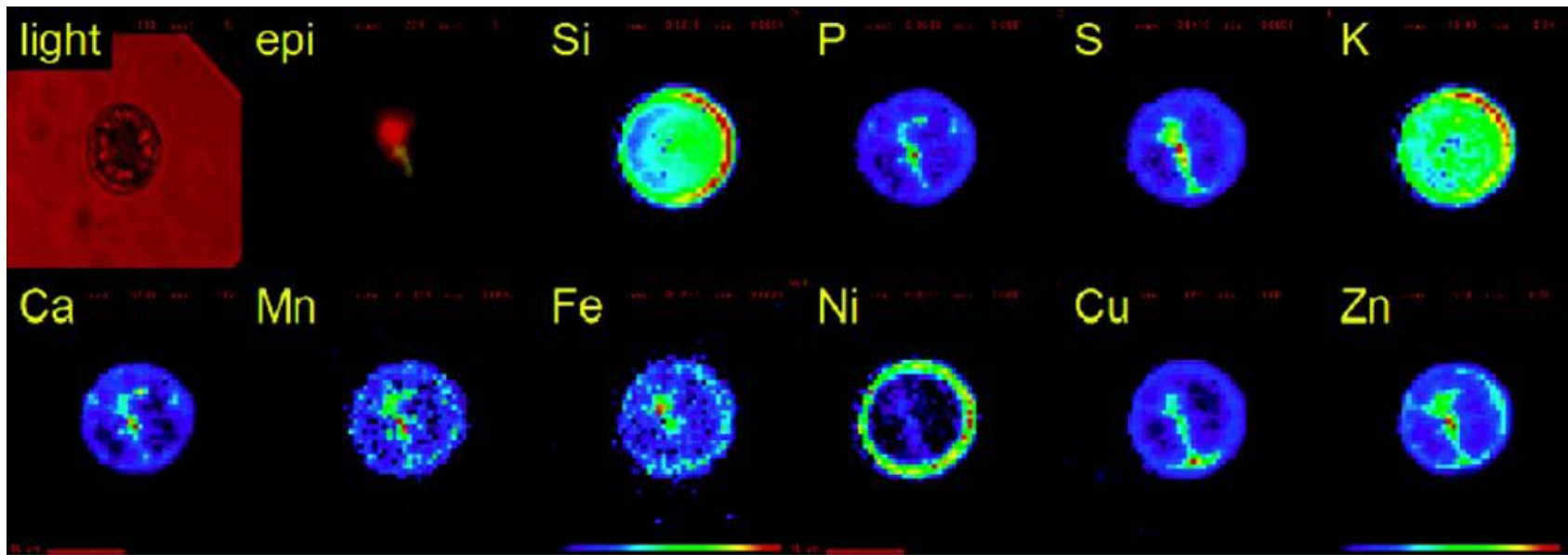
Pennate diatoms – initial cells produced by non-motile gametes

# Diatoms from the Gulf of Alaska





Girdle bands



Using synchrotron x-ray fluorescence to map out elements in a diatom cell (Zhao & Le, 2007)

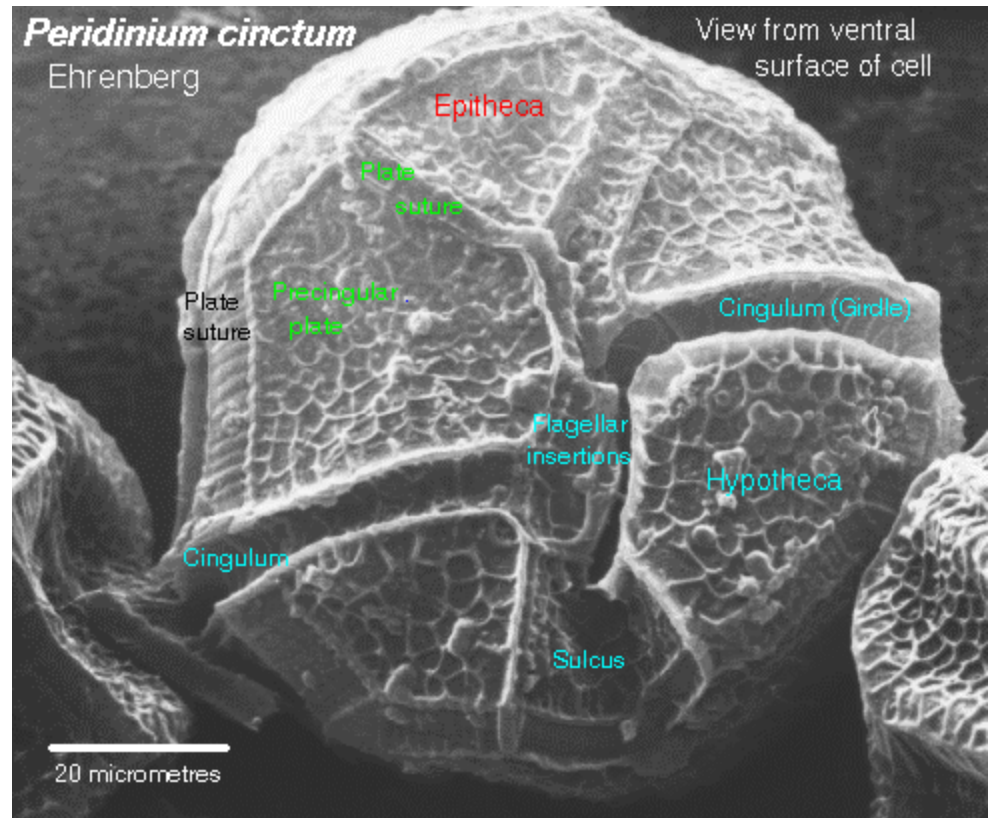


# Dinophyceae

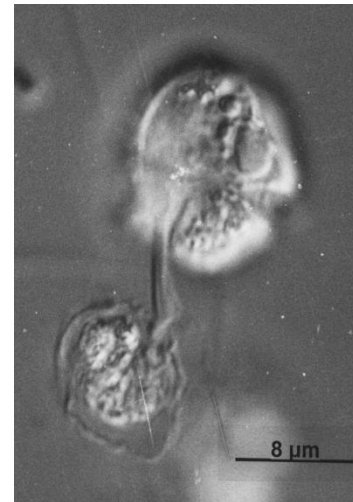
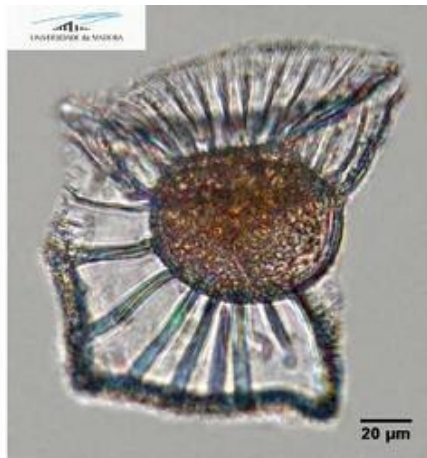
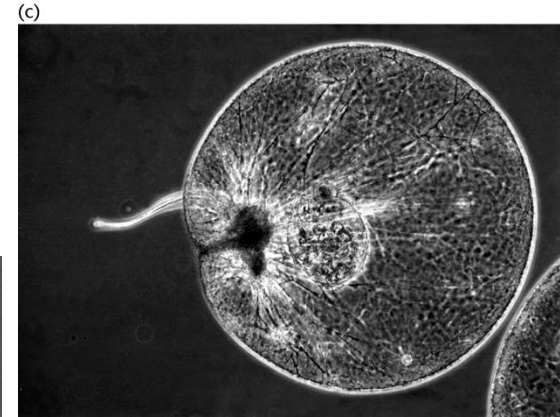
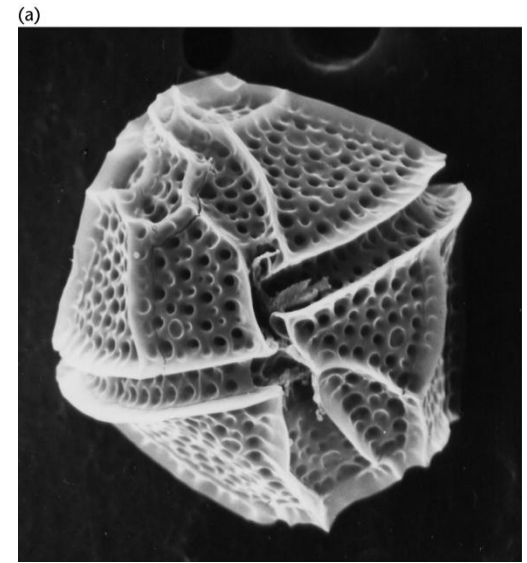
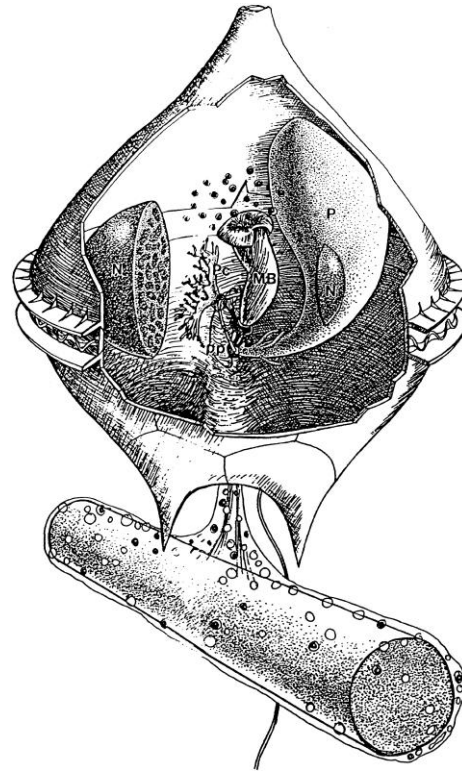
- Most unicellular, some colonial
- Complex group, including autotrophs, heterotrophs, and mixotrophs
- Two dissimilar flagella; one is transverse ('side-to-side') and one is longitudinal ('up-down') each bearing fine lateral hairs; both arise on the ventral side of the cell
- Transverse furrow and longitudinal groove present in most taxa
- Chloroplasts surrounded by 3 membranes
- Chl  $a$ , chl  $c_2$ ; accessory pigments are peridinin,  $\beta$ -carotene, and other xanthophylls
- Pyrenoids present
- Reserve polysaccharide is starch
- Cell wall made of cellulose arranged in plates (thecal plates) – pellicle made up of cell membrane externally and underlain by flattened vesicles (plates)
- Many have eyespots
- Possess trichocysts (discharge explosively when simulated, throwing out transversely striated, four-sided threads)

## Dinoflagellates

- theca (robust 'envelope')
- epitheca (=epicone)
- hypotheca (= hypocone)
- transverse groove (cingulum or girdle)
- longitudinal groove (sulcus)
- dino = 'spinning' – swim in a turning motion
- many produce toxins and are considered to be harmful algal bloom species

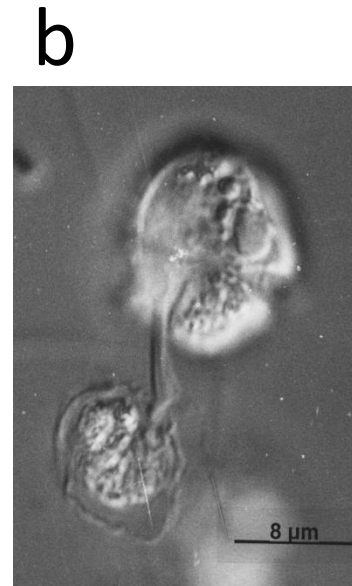
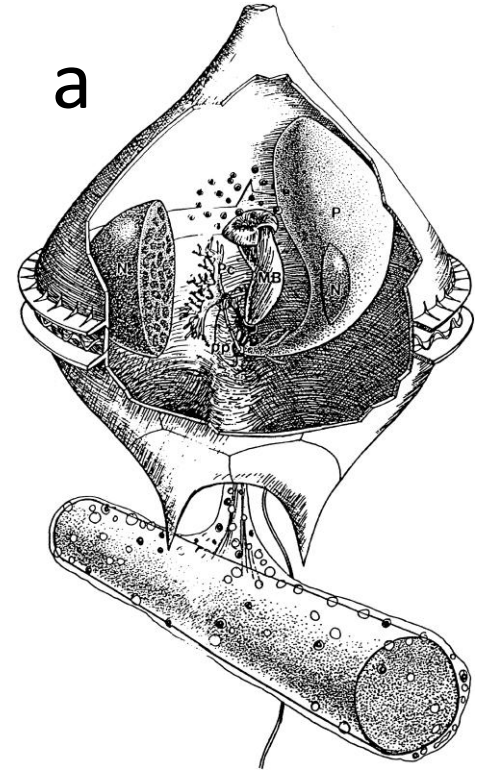


# Dinophyceae



# Dinophyceae as predators

- Some dinoflagellates prey on other organisms using an extracellular net of fibers or mucus-like material (a pallium); extruded through pallial pore located in the sulcus; digestion occurs in the pallial sac (a)
- Another strategy is to insert a peduncle into the tissue of the prey (b)



# Chrysophyceae

- Unicellular or colonial
- May or may not be flagellate
- Flagella inserted near the apex of the cell (not laterally)
- Golden-brown chloroplasts (chl is masked by the accessory pigment, fucoxanthin); may also possess zeaxanthin, antheraxanthin, violaxanthin, diatoxanthin, diadinoxanthin
- Some species bear siliceous scales
- Freshwater and marine
- Chrysolaminarin as storage product
- Common in oligotrophic lakes; greater diversity in freshwater



[www.conncoll.edu](http://www.conncoll.edu)



<http://www.members.shaw.ca>

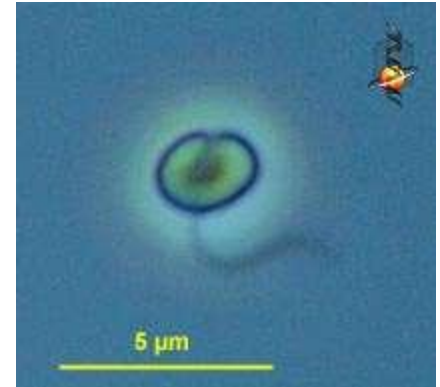


# Pelagophyceae

- Small, flagellated cells
- Some form 'brown tides' (e.g. *Aureococcus anophagefferens*)
- Coastal and open ocean environments



[www.bayshorewatershed.org](http://www.bayshorewatershed.org)

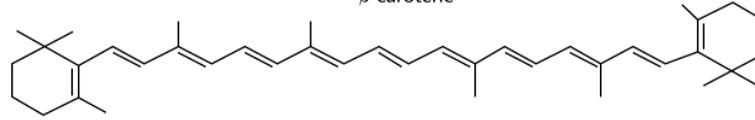


*Encyclopedia of Life*

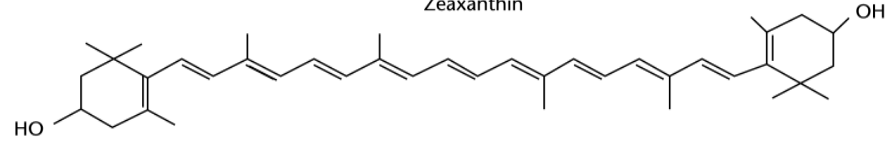


(b)

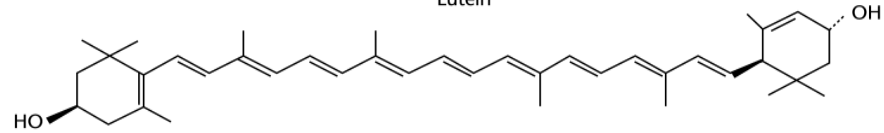
$\beta$ -carotene



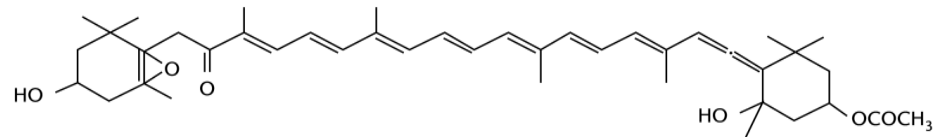
Zeaxanthin



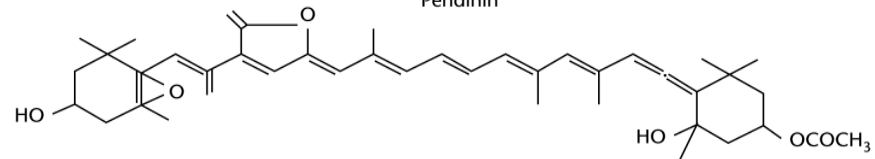
Lutein



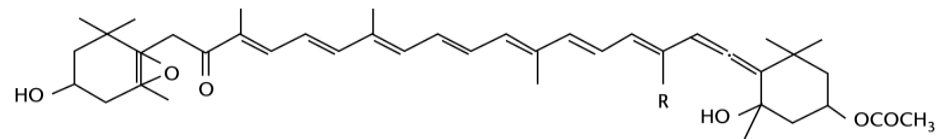
Fucoxanthin



Peridinin



Fucoxanthin derivatives

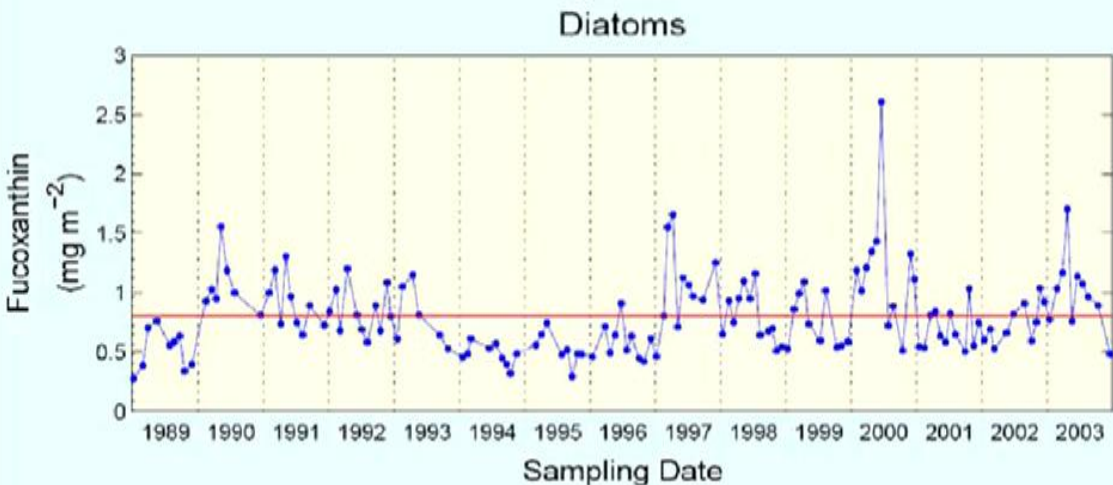
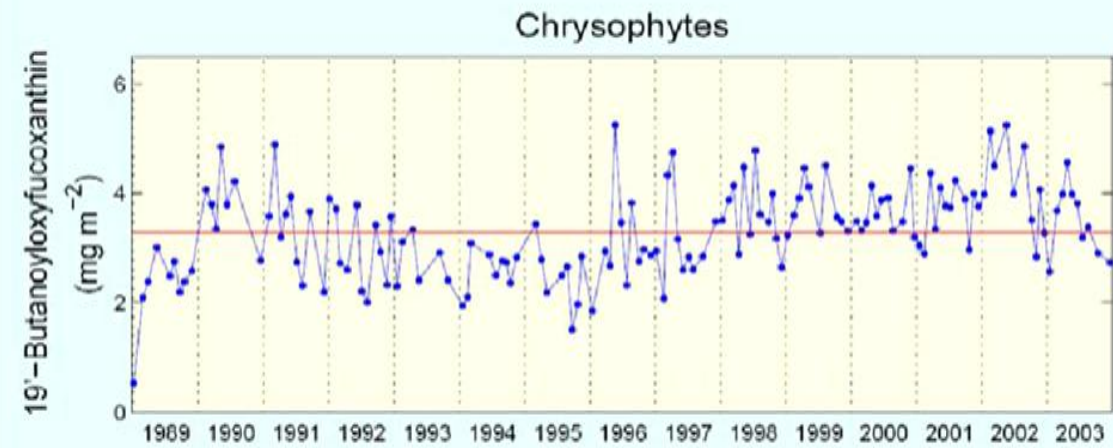


19'-hexanoyloxy-fucoxanthin, R = CH<sub>2</sub> — O — C — C<sub>3</sub>H<sub>7</sub>

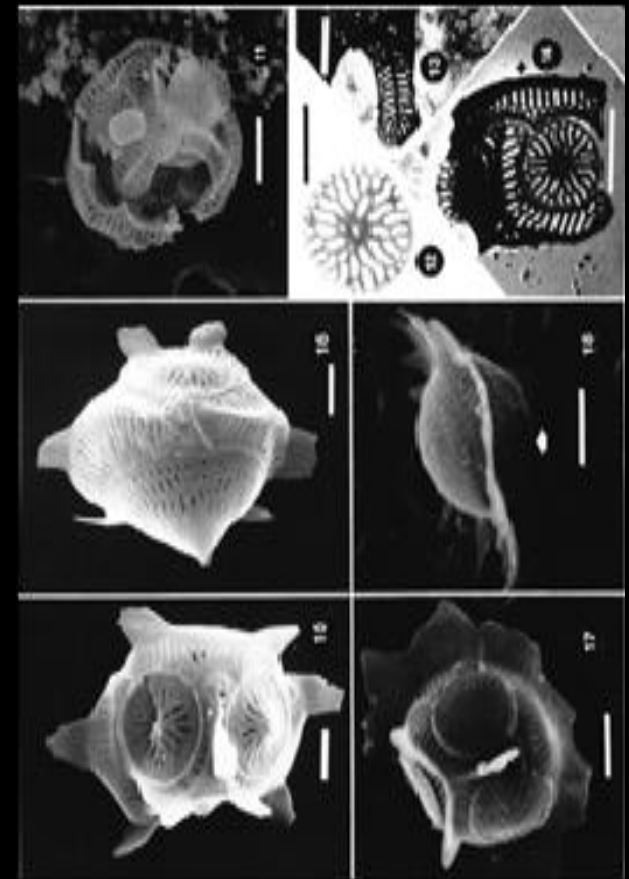
19'-butanoyloxy-fucoxanthin, R = CH<sub>2</sub> — O — C — C<sub>3</sub>H<sub>11</sub>

Class	Pigments	Diagnostic pigment
Bacillariophyceae	Chl <i>a</i> , chl <i>c</i> <sub>2</sub> , fucoxanthin, diadinoxanthin, β-carotene, diatoxanthin, chl <i>c</i> <sub>1</sub>	Fucoxanthin
Chlorophyceae	Chl <i>a</i> , chl <i>b</i> , β-carotene, lutein, violaxanthin, zeaxanthin, neoxanthin	Chl <i>b</i>
Chrysophyceae	Chl <i>a</i> , chl <i>c</i> <sub>1</sub> , <i>c</i> <sub>2</sub> , fucoxanthin, 19'butanoyloxyfucoxanthin, diadinoxanthin	Chl <i>c</i> <sub>1</sub> , <i>c</i> <sub>2</sub>
Cryptophyceae	Chl <i>a</i> , chl <i>c</i> <sub>2</sub> , β-carotene, alloxanthin, phycoerythrin & phycoerythrin (cryptophyte types)	Alloxanthin
Cyanophyceae	Chl <i>a</i> , β-carotene, zeaxanthin, phycocyanin, allophycocyanin, phycoerythrin	Zeaxanthin
Prochlorophyceae	Chl <i>a</i> , divinyl chl <i>b</i> , β-carotene, zeaxanthin	Divinyl chl <i>b</i>

Class	Pigments	Diagnostic pigment
Dinophyceae	Chl <i>a</i> , <i>c</i> <sub>2</sub> , β-carotene, peridinin, dinoxanthin, diadinoxanthin	Peridinin
Haptophyceae	Chl <i>a</i> , chl <i>c</i> <sub>1</sub> or <i>c</i> <sub>3</sub> , chl <i>c</i> <sub>2</sub> , β-carotene, fucoxanthin, 19'hexanoyloxyfucoxanthin, 19'but, diadinoxanthin, diatoxanthin	19' hexanoyloxyfucoxanthin
Prasinophyceae	Chl <i>a</i> , chl <i>b</i> , prasinoxanthin	Prasinoxanthin
Pelagophyceae	Chl <i>a</i> , chl <i>c</i> <sub>2</sub> , chl <i>c</i> <sub>3</sub> , diatoxanthin, fucoxanthin, β-carotene, diadinoxanthin, 19'butanoyloxyfucoxanthin	19'butanoyloxyfucoxanthin
Euglenophyceae	Chl <i>a</i> , chl <i>b</i> , β-carotene, diadinoxanthin, zeaxanthin, neoxanthin	Violaxanthin, chl <i>b</i>
Raphidophytes	Chl <i>a</i> , <i>c</i> <sub>1</sub> , <i>c</i> <sub>2</sub> , fucoxanthin, β-carotene, violaxanthin	



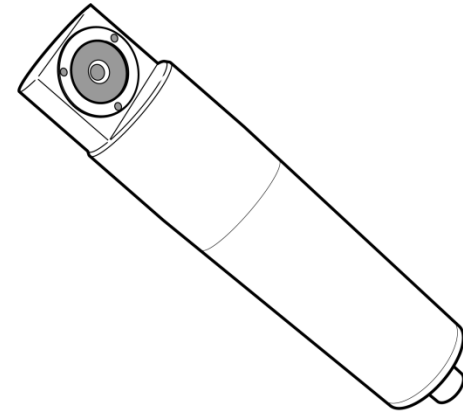
The various groups (or 'taxa') respond to changes in the environment in different ways



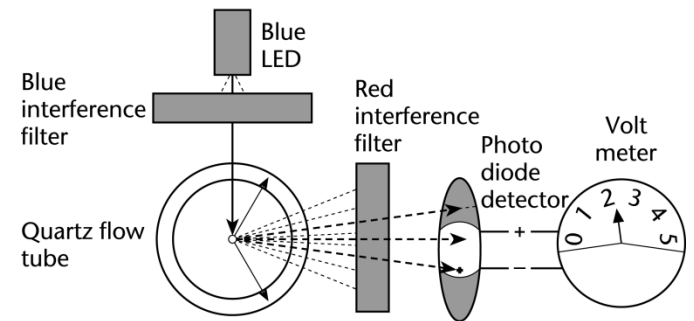
*Letelier, 2006*

# Measuring phytoplankton

- Use chlorophyll *a* as a proxy
  - Spectrophotometer (absorption)
  - Fluorometer (fluorescence)
- Note: artifactual day-night and depth variation
  - fluorescence varies strongly with external illumination
  - chlorophyll decouples in the dark from the energy transfer system in the chloroplasts of the plant and fluoresces more strongly

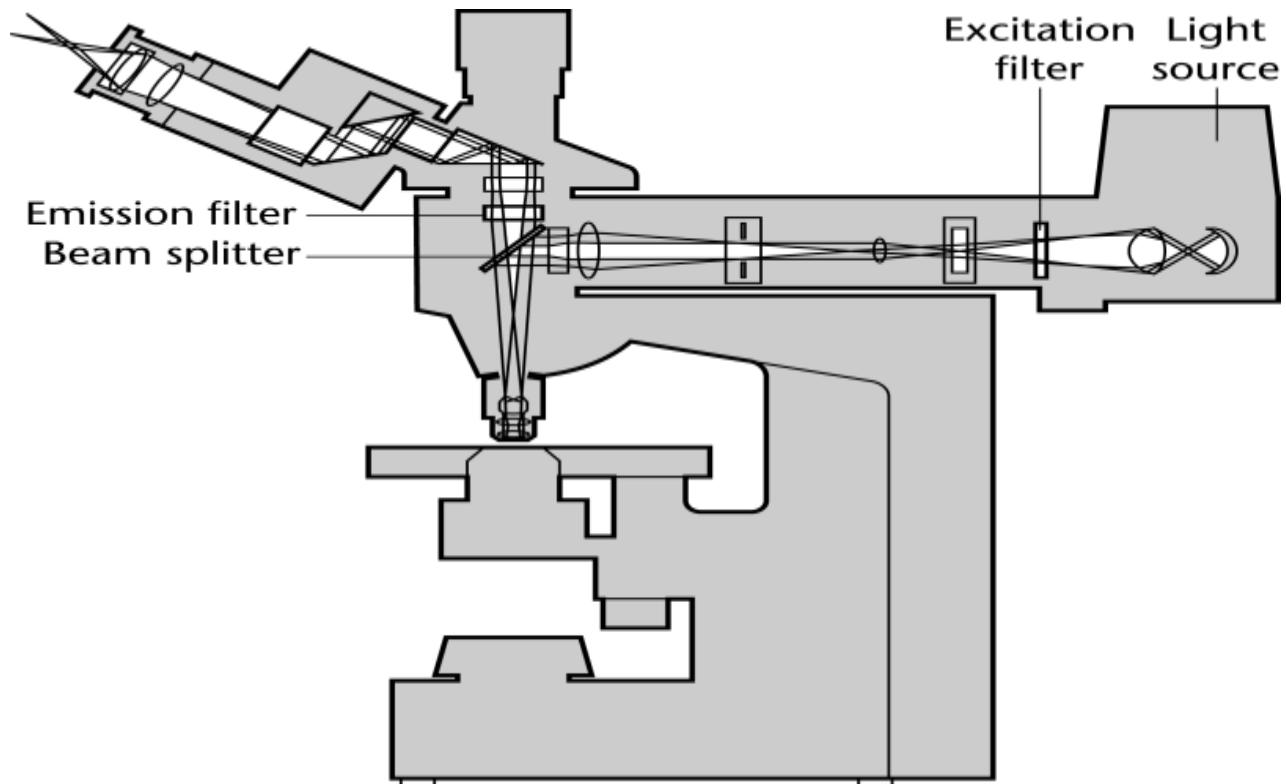


**Theory of operation**



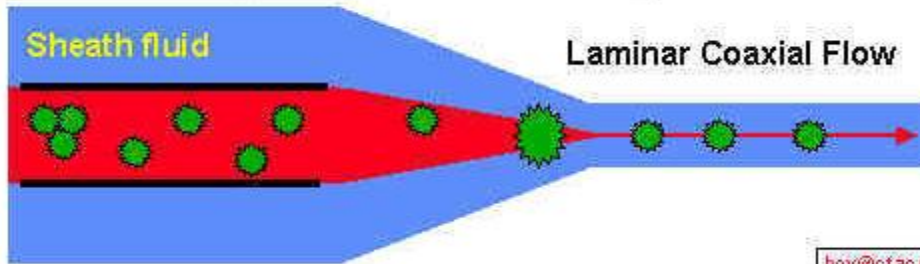
- Phytoplankton
- Blue (455 nm) excitation light (incident and scattered)
- > Red (685 nm) emission light

# Epifluorescence microscope

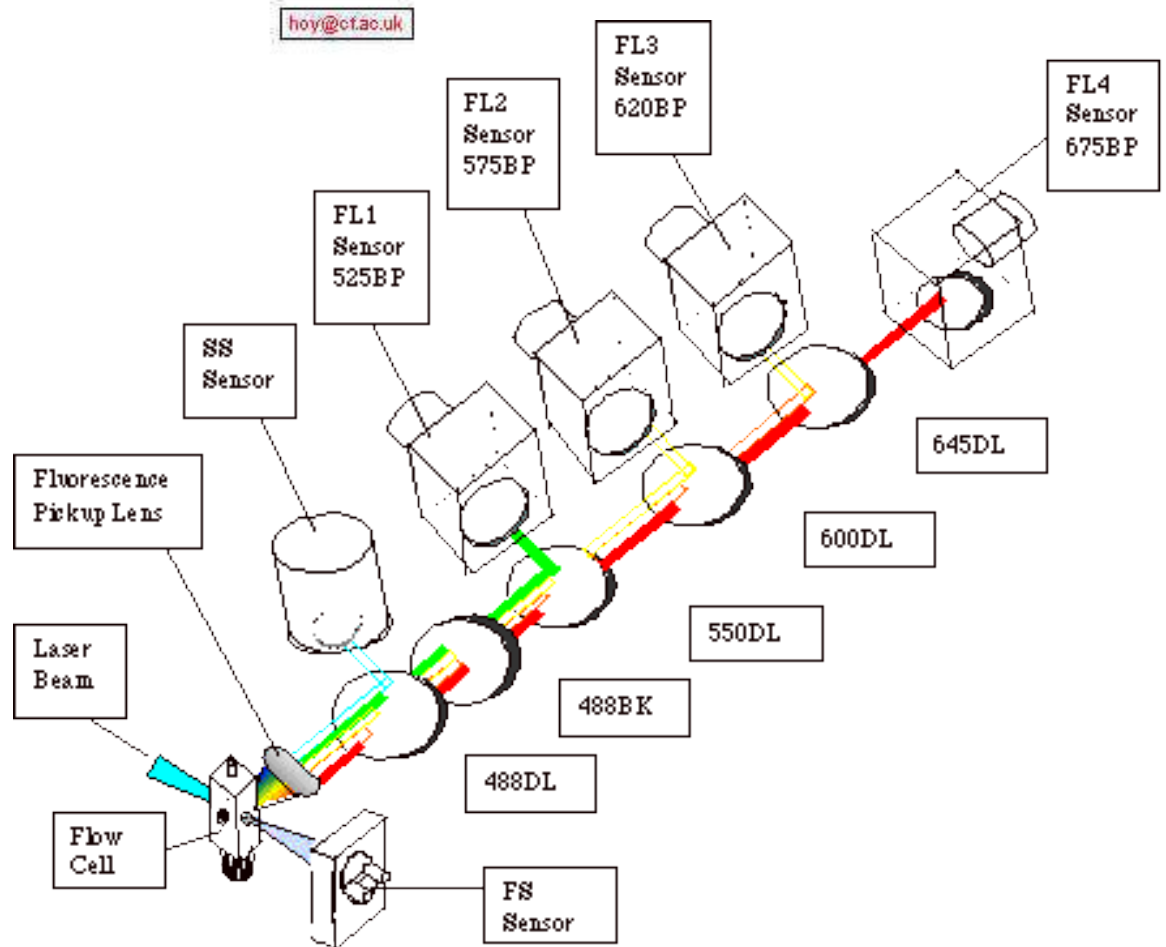




# Hydrodynamic Focusing



# Flow cytometer

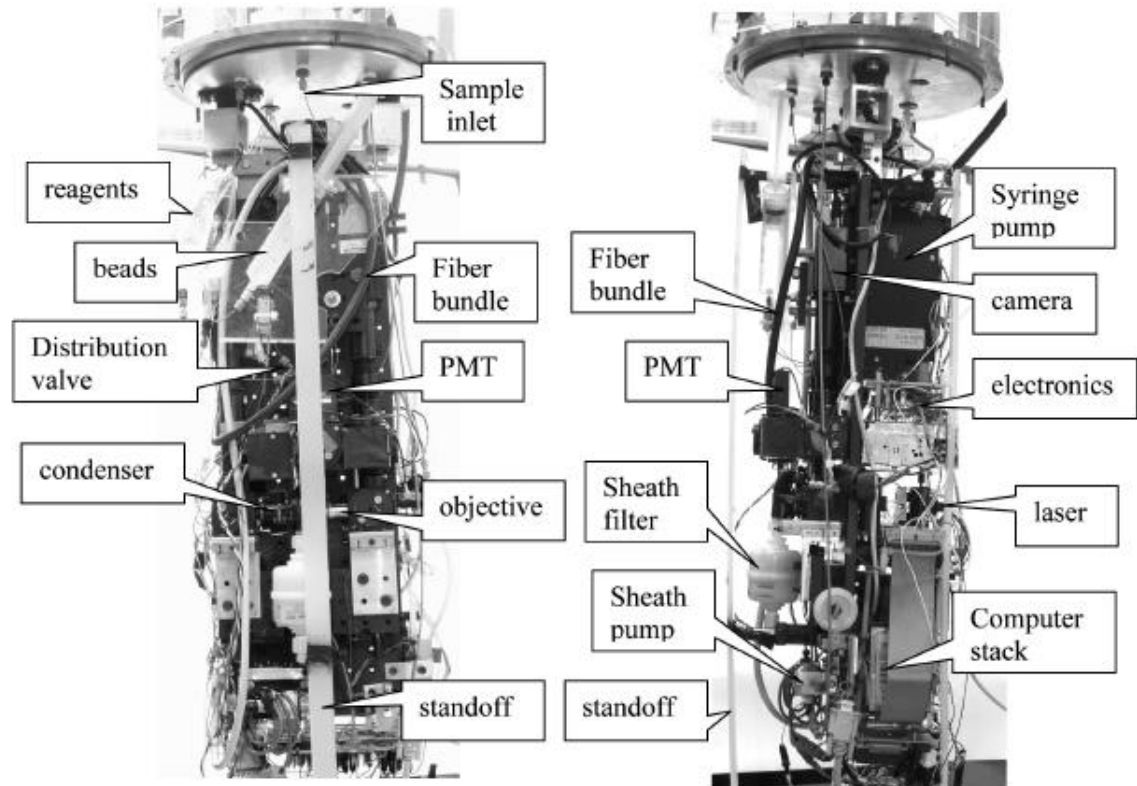


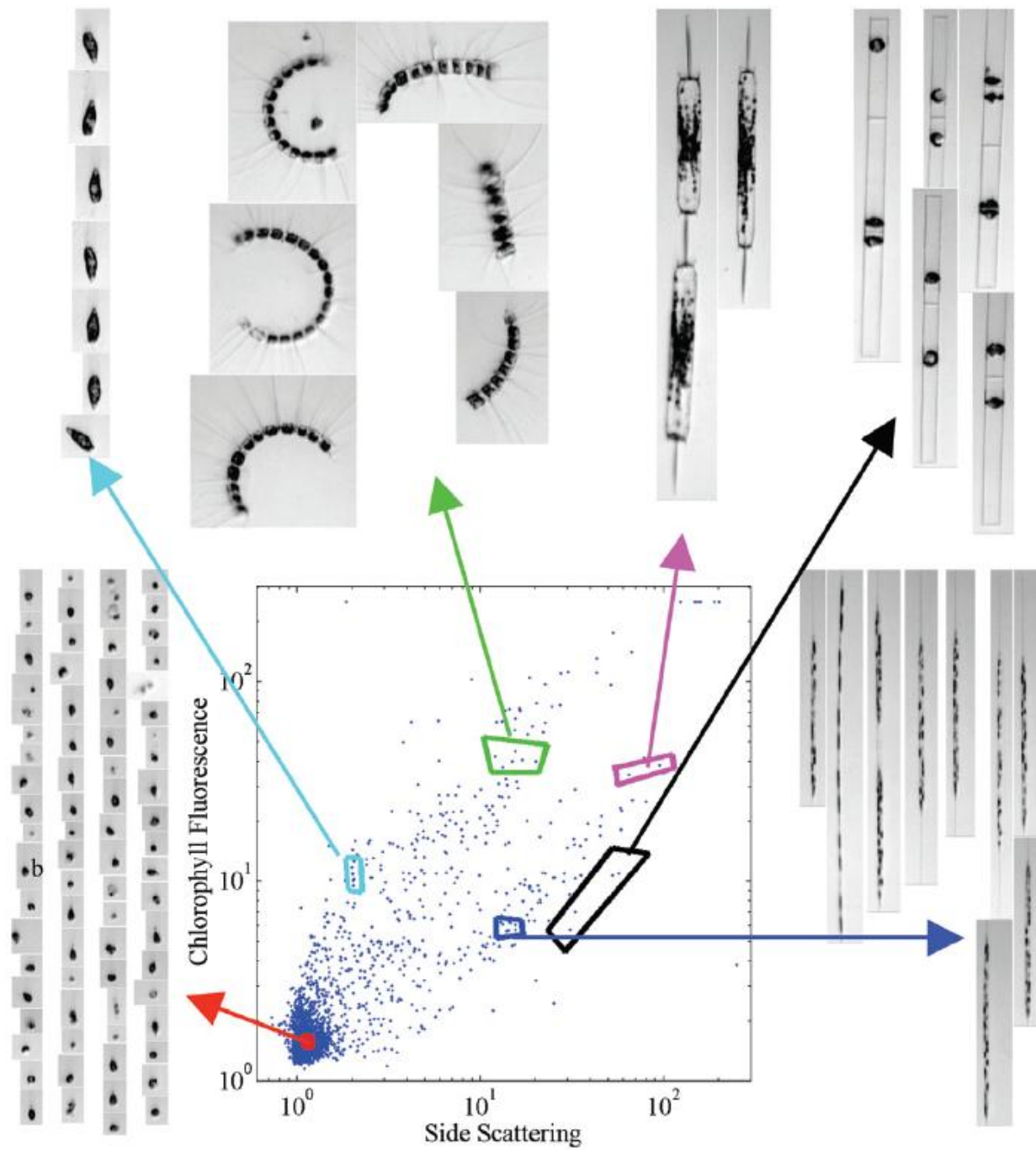
Olson and Sosik, Woods  
Hole Oceanographic  
Institute



Cytobuoy

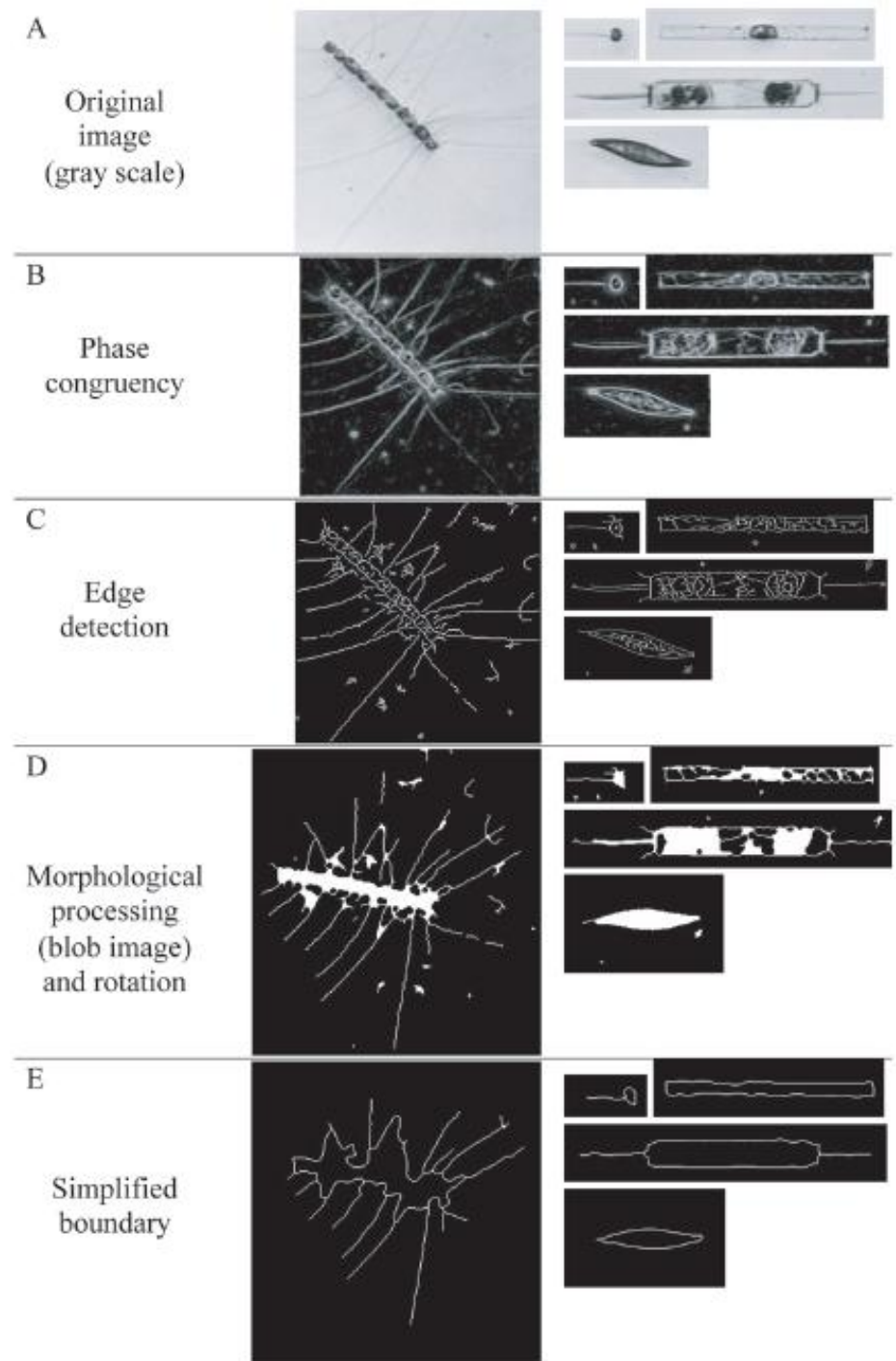
FlowCytobot  
and Imaging  
FlowCytobot



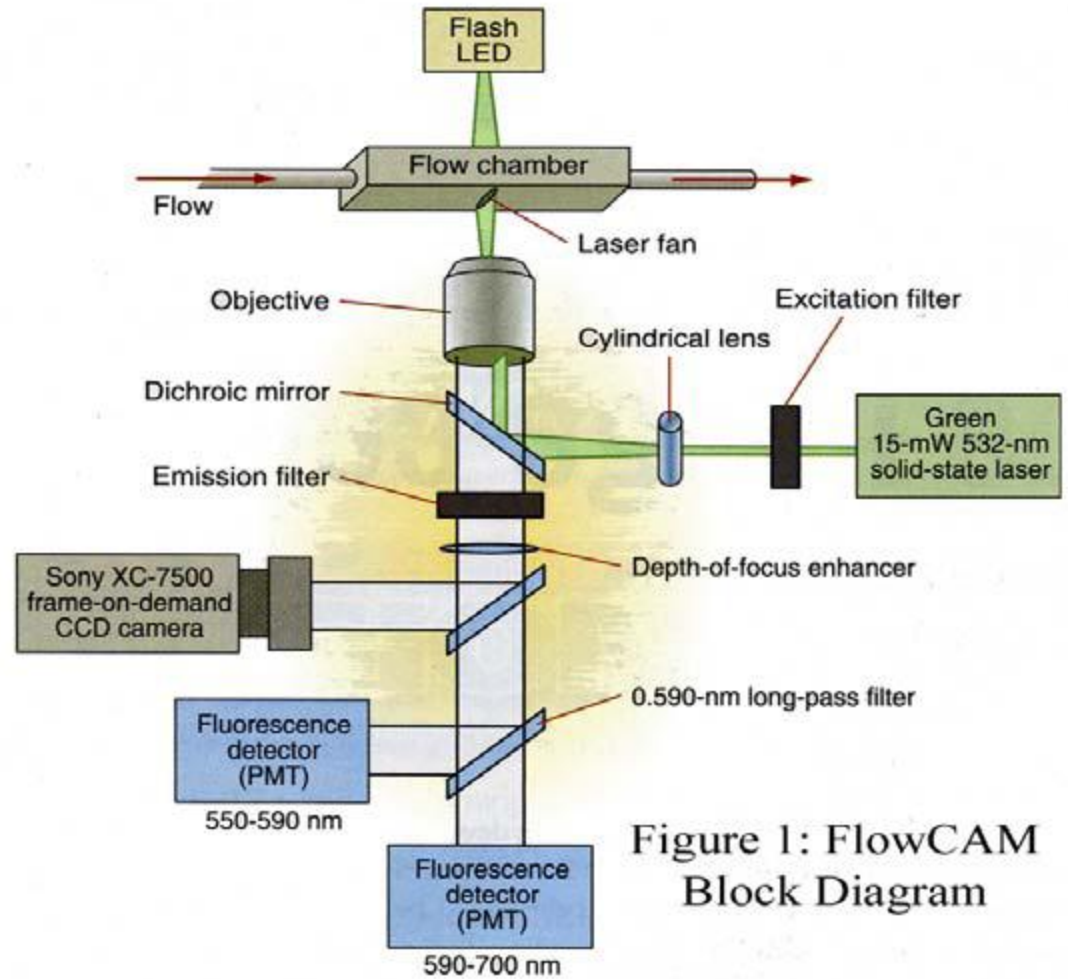


# Image processing steps for Flow CytoBot

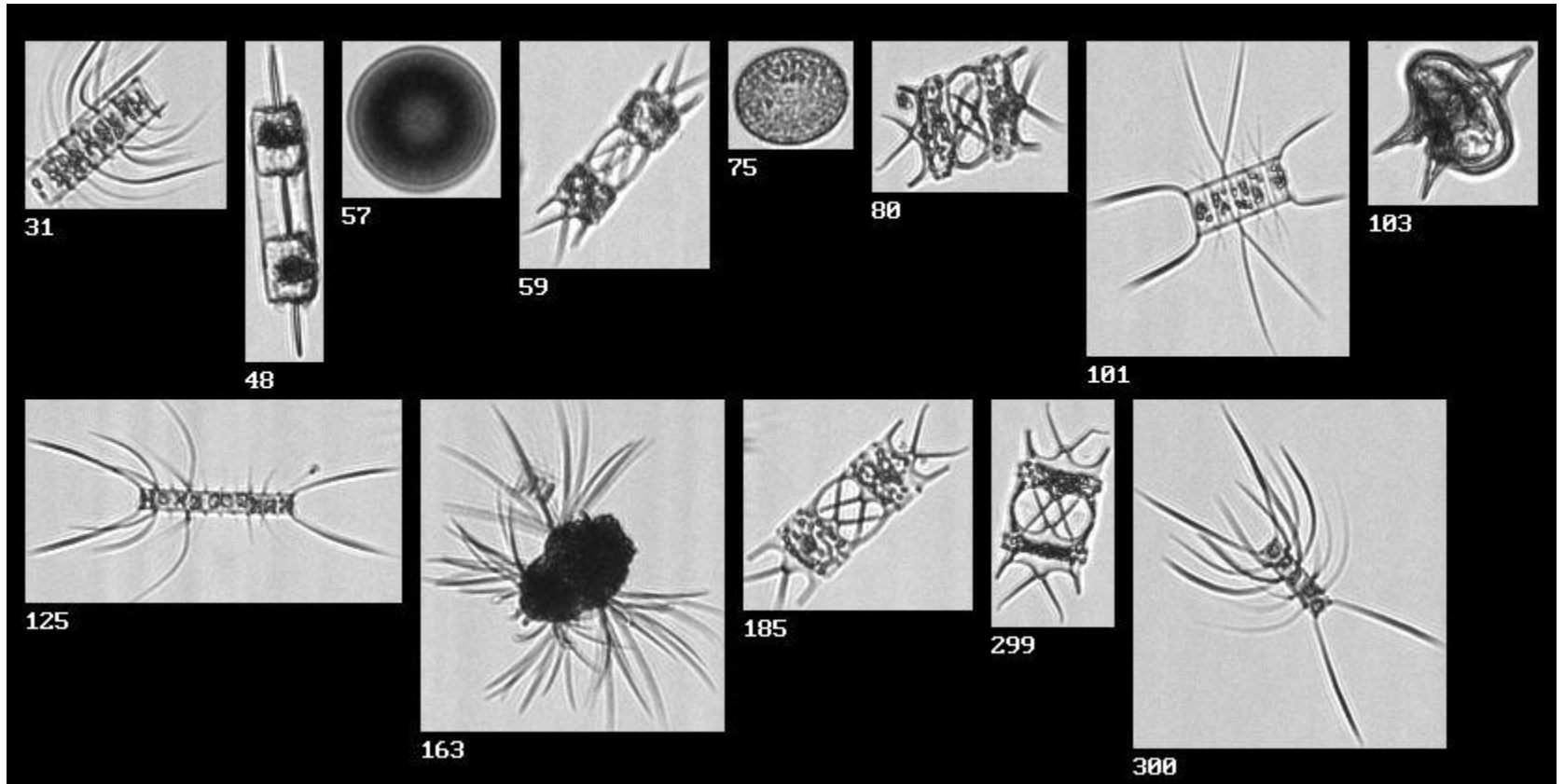
Sosik & Olsen, 2007



# FlowCAM: Flow Cytometer And Microscope



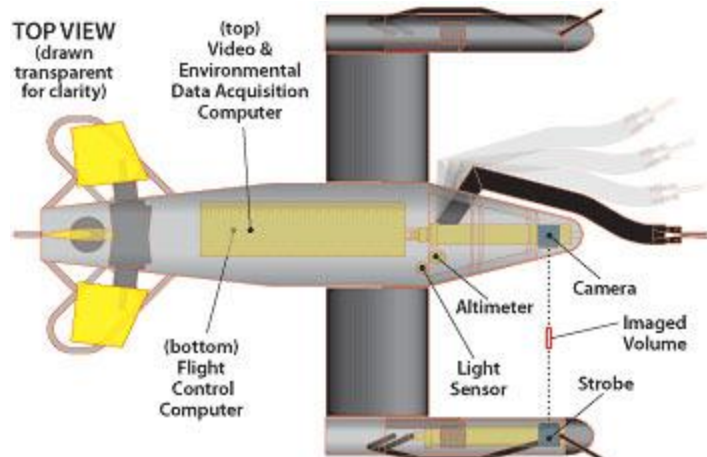
# Marine sample from the coast of Oregon



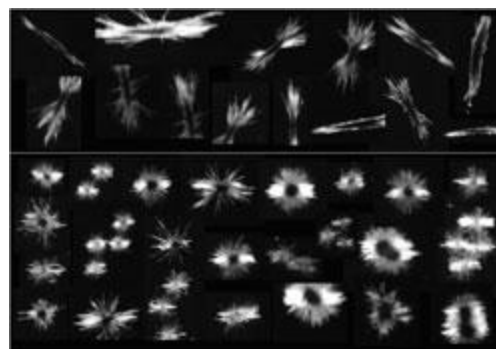
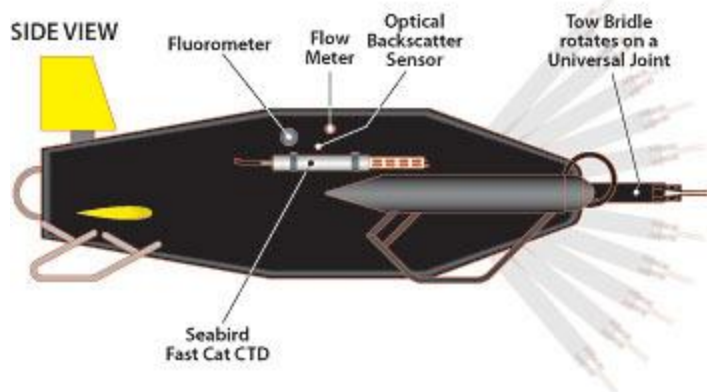




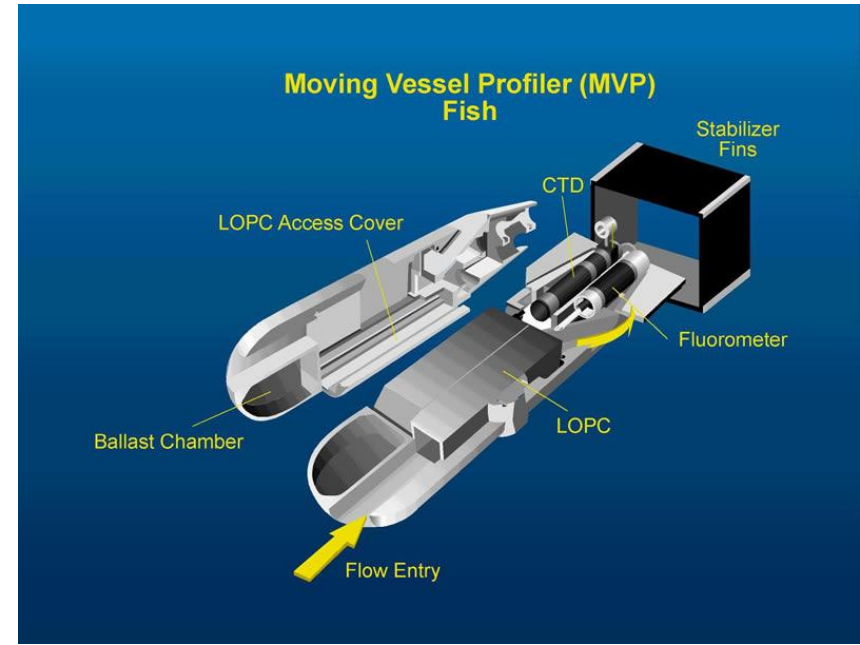
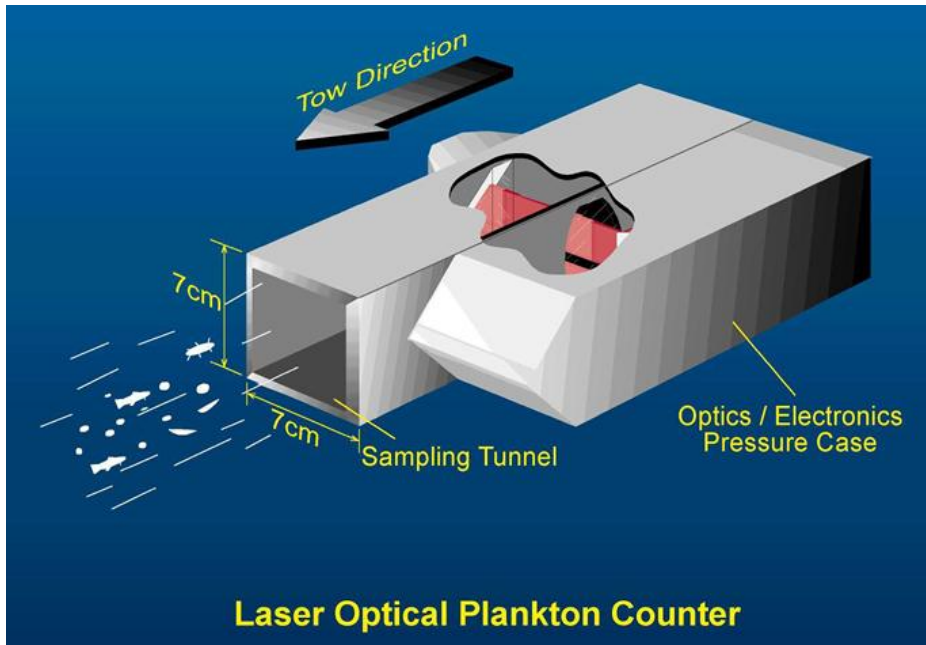
TOP VIEW  
(drawn transparent for clarity)



SIDE VIEW



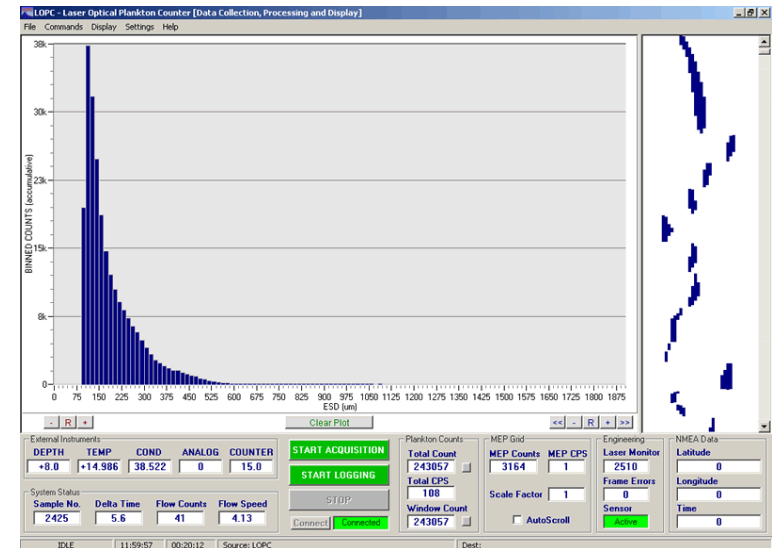
# Video Plankton Recorder



( A ) Wide Tunnel LOPC - Batfish Tow



Shape profiles obtained from an LOPC mounted on a Batfish vehicle towed at 8 knots. Silhouettes show profiles of euphausiids and copepods of a variety of sizes. Credit: ©Alex Herman, Bedford Institute of Oceanography.

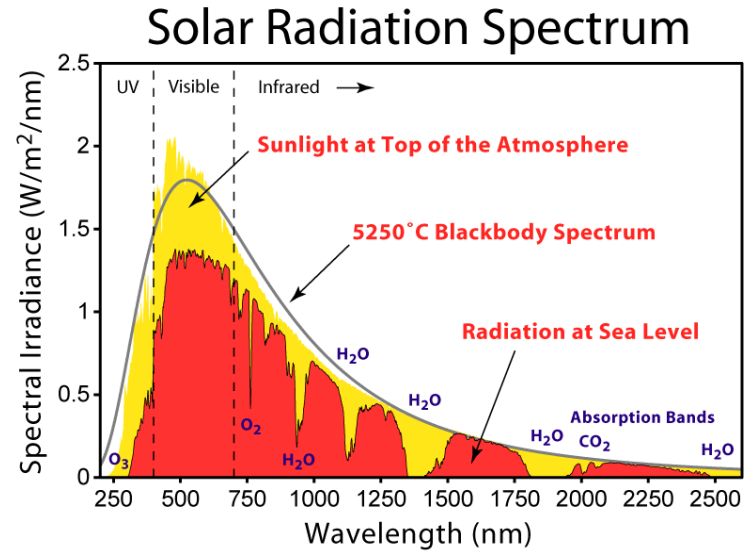
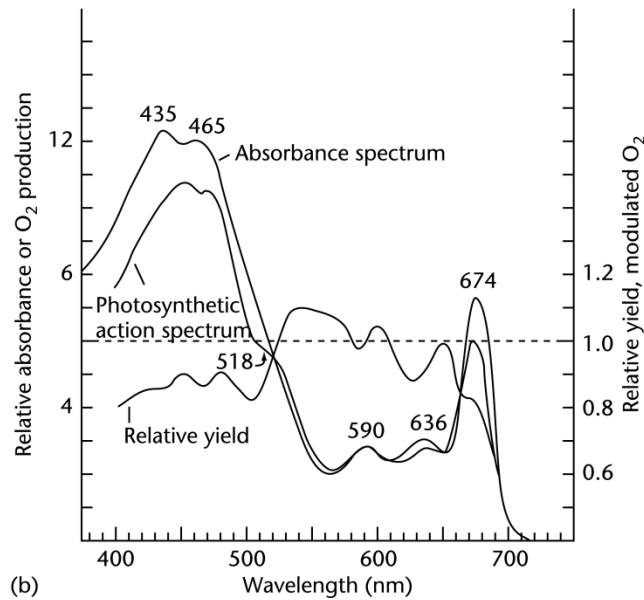
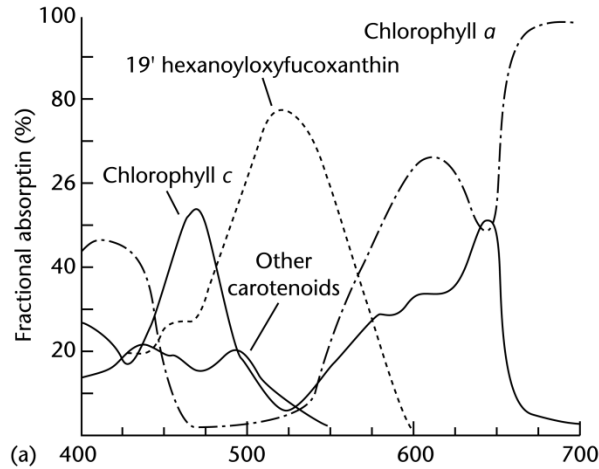


# Primary production

- Fixation of inorganic carbon using energy from the sun to make organic carbon (through the process of photosynthesis)
- **Primary production** = organic matter produced (speak of a *rate* of primary production)
- Primary productivity = rate of organic matter production – never say ‘rate’ of primary productivity
- Generally measured using a tracer of carbon uptake ( $H^{14}CO_3^-$  or  $H^{13}CO_3^-$ ) or by appearance of oxygen (since oxygen evolution is the product of photosynthesis)
- Geochemical ways: apparent oxygen utilization, etc.
- **New production**: Dugdale, 1967
  - Ratio of uptake of nitrogen species used tells us about what is getting exported in a steady-state system (‘f ratio’)
  - ‘new’ nitrogen is nitrate (and  $N_2$  gas), while regenerated nitrogen is ammonium and organic nitrogen (e.g. urea)
  - This concept is losing favor as we discover many intricacies in the nitrogen cycle

$$f \text{ ratio} = \frac{\rho NO_3^-}{\rho NO_3^- + \rho NH_4^+}$$

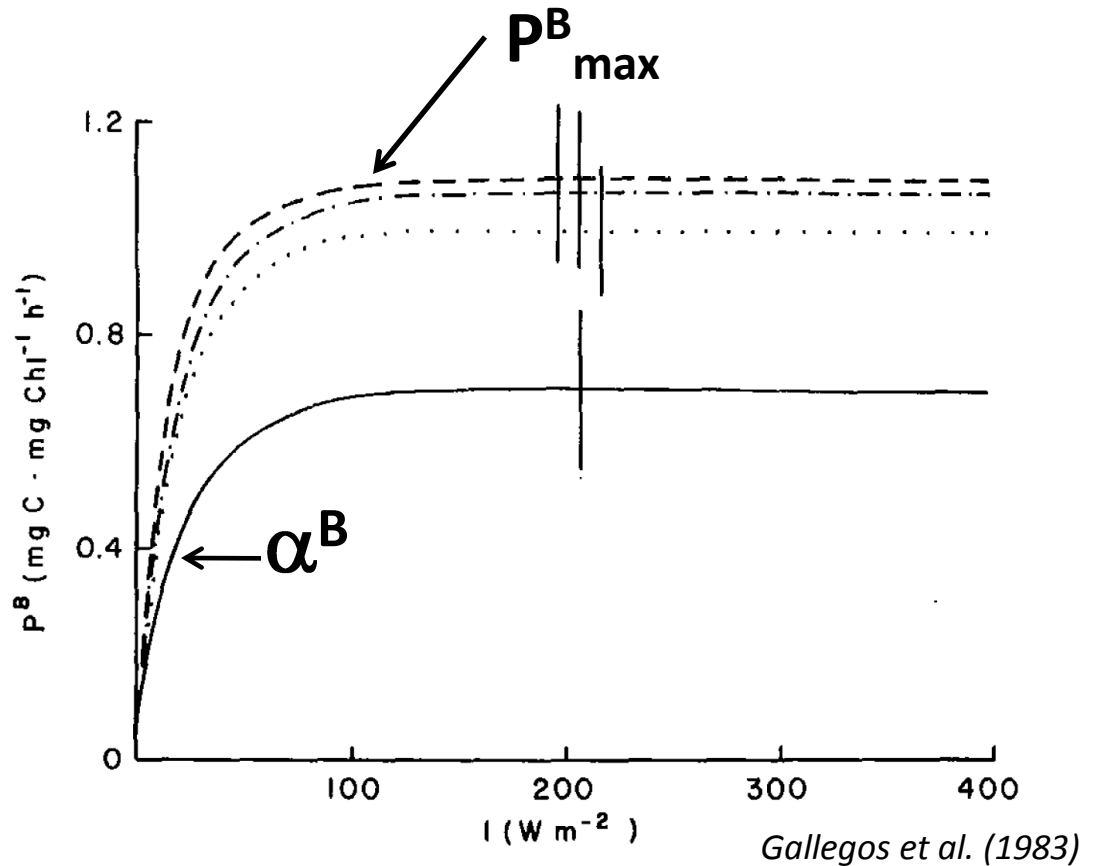
# There is a correspondence between spectra of pigment absorption and solar radiation



# Photosynthesis vs. irradiance (P vs. E) curves

- Alpha ( $\alpha$ ): initial slope of the PE curve
- $P_{max}^B$ : maximum chlorophyll-specific rate of C-fixation

Chlorophyll-specific carbon fixation



Beta ( $\beta$ )

Irradiance ( $\text{W m}^{-2}$  or  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ )

# Photoinhibition

- $\beta$ : slope of the PE curve that reflects photoinhibition
- Photoinhibition: light-induced reduction in photosynthetic capacity caused by damage to photosystems (mainly PS-II)

